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Remarks:

The applicant has subsequently filed a sequence listing and declared, that it includes no new matter.

(54) Cellulose synthase gene

(57) mRNA was extracted at the stage for cotton plant fibrous cells to accumulate cellulose, and cDNA's complementary thereto were synthesized to construct a cDNA library. Clones of a number of 750 were arbitrarily selected from the library, and they were randomly subjected from to sequencing. Those having homology to

an amino acid sequence deduced from a gene of cellulose 4- β -glucosyltransferase (bcsA) of cellulose synthase operon of acetic acid bacterium were selected from obtained nucleotide sequences of the respective clones. Thus, DNA coding for cellulose synthase was obtained.

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DescriptionTechnical Field

5 The present invention relates to a DNA coding for cellulose synthase originating from cotton plant (Gossypium hirsutum), a recombinant DNA containing the same, a transformed cell transformed with the DNA, and a method for controlling cellular cellulose synthesis.

Background Art

10 Cellulose is used for paper, woody structural materials, fiber, cloths, food, cosmetics, and pharmaceuticals, as well as it is utilized as energy. Therefore, cellulose is industrially useful and valuable. Cellulose is capable of forming a variety of crystalline structures, and hence it is expected to develop a new material by controlling enzymes involved in biosynthesis of cellulose. The cellulose-related industry has been hitherto directed to such cellulose products that
 15 have been already produced, in which there has been no trial to develop a new material based on an aspect of biosynthesis. The mechanism of disease action, which is exerted by pathogenic microorganisms on plants, often results from the inhibition on cellulose biosynthesis as in Pyricularia oryzae (P. oryzae). Therefore, the addition of disease resistance to the cellulose biosynthesis mechanism is agriculturally applicable and valuable. Further, cellulose is the most abundant organic compound on the earth, and it is a sink in which the largest amount of CO₂ in the atmospheric
 20 air is fixed. Therefore, the genetic improvement of cellulose biosynthesis enzymes is also applicable to the industry which is directed to the control of CO₂ in the atmospheric air based on the use of cellulose as the sink.

In recent years, cDNA's originating from fiber cells of cotton plant have been randomly sequenced, and it has been reported that full length CelsA1 and partial length of CelsA2 probably represent cDNAs of cotton plant cellulose synthase, in view of the homology to bacterial cellulose synthase gene (bacterial BcsA) (Pear et al., Proceeding of National
 25 Academy of Science, USA (1996) 93 12637-12642). The binding ability to UDP-glucose has been demonstrated for CelsA1. However, as for CelsA2, the homology has been merely demonstrated for the C-terminal amino acid sequence.

Disclosure of the Invention

30 The present invention has been made in order to provide a new method for regulating cellulose production in prokaryotic cells or eukaryotic cells, an object of which is to provide a DNA coding for cellulose synthase, a recombinant DNA containing the same, a transformed cell transformed with the DNA, and a method for regulating cellular cellulose synthesis.

The present inventors firstly extracted mRNAs at the stage for cotton plant fiber cells to accumulate cellulose, and
 35 cDNAs complementary thereto were synthesized to construct a cDNA library. 750 of cDNA clones were arbitrarily selected from the library, and they were randomly subjected to sequencing. Six amino acid sequences were derived for one nucleotide sequence of each of the obtained clones to select those having homology to an amino acid sequence obtained by translation from a gene of cellulose 4-β-glucosyltransferase (bcsA) of cellulose synthase operon of acetobacterium. As a result, genes, which were classified into three types or groups, were found, and they were designated
 40 as PcsA1, PcsA2, and PcsA3 respectively (PcsA is an abbreviation of "Plant Cellulose Synthase A").

That is, the present invention lies in a DNA coding for any one of the following proteins (A) to (C):

- 45 (A) a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO: 2 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 2;
- (B) a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO: 4 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 4; and
- 50 (C) a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO: 8 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 8, and comprising an amino acid sequence shown in SEQ ID NO: 11 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 11.

55 In another aspect, the present invention provides a recombinant vector comprising all or a part of the DNA as defined above, and a transformed cell transformed with the DNA as defined above.

In still another aspect, the present invention provides a method for regulating cellulose synthesis in a cell, comprising the steps of introducing the DNA as defined above into the cell, and expressing RNA having a nucleotide

sequence homologous to the DNA as defined above or a nucleotide sequence complementary to the DNA as defined above.

SEQ ID NO: 1 corresponds to a sequence of PcsA1, and SEQ ID NO: 3 corresponds to a sequence of PcsA2. SEQ ID NO: 5 corresponds to a sequence of 3'-side region of PcsA3, SEQ ID NO: 7 corresponds to a sequence of 5'-side region of PcsA3, and SEQ ID NO: 9 corresponds to a sequence of internal region of PcsA3.

It has been demonstrated that PcsA1 and PcsA2 of the DNA's described above are DNA's coding for cotton plant cellulose synthase, according to the expression in eukaryotic cells (animal cells and/or yeast). It has been also demonstrated that an antibody thereagainst inhibits the cotton plant cellulose synthase activity in a cell-free system. Further, PcsA3, which is different from PcsA1 and PcsA2, has been found. Any one of these species was obtained as partial one, at the stage of clones obtained by the random sequencing, and no 5'-portion of the coding region was contained. Therefore, clones which have sequences of 5'-portions were isolated in accordance with the 5'-RACE method based on the use of PCR to determine the sequences. As a result of this operation, the sequences of the 5'-portions corresponding to the partial length clones were obtained for PcsA1 and PcsA2.

On the other hand, as for PcsA3, a sequence of a 5'-portion of another clone, which was considered to belong to the same PcsA3 group, was obtained. The both sequences had extremely high homology, and hence they were considered to have underwent multiple gene formation relatively recently originating from an identical gene through the process of duplication. Therefore, even when the both are combined with each other at corresponding portions to construct a fused gene followed by expression, it is assumed that the activity and function of a produced enzyme may not be affected thereby.

As for PcsA1 and PcsA2, in order to obtain a full length clone, primers were designed on the basis of the sequence of the 5'-portion and the sequence of the 3'-portion of the partial length clone to perform PCR. Thus, a clone containing ORF was obtained.

Those applicable as the template to be used for the RACE method may be any of cDNA synthesized from mRNA and a phage library. When the phage library is used, it is possible to use a sequence in the vector as a 5'-side primer.

As a result of random sequencing, seven clones concerning PcsA2 were most abundantly present, of 15 clones seemed to code the cellulose synthase. Expression was confirmed in eukaryotic cells (animal cells and/or yeast) transformed with the cellulose synthase gene. As a result, the cellulose synthase activity was observed.

The present invention will be explained in detail below.

<1> Preparation of cotton plant cDNA library

Cotton plant fiber cells at the stage of cellulose accumulation are preferably used as a material for extracting mRNA to construct a cotton plant cDNA library. The method for extracting mRNA is not specifically limited, for which it is possible to adopt an ordinary method for extracting mRNA from plant.

cDNA can be synthesized, for example, by using a poly T sequence which is complementary to poly A nucleotide existing at the terminal of mRNA as a primer to synthesize complementary DNA by the aid of reverse transcriptase, and forming a double strand by the aid of DNA polymerase.

The method therefor is described, for example, in Molecular Cloning (Maniatis et al., Cold Spring Harbour Laboratory). However, a variety of cDNA synthesis kits are commercially available from various companies, which may be used.

Generally, the library is constructed by using a phage vector. A variety of commercially available vectors are usable. However, it is preferable to use a vector, for example, λ ZAP vector in which it is unnecessary to perform recloning from the vector, and it is possible to immediately prepare a plasmid for sequencing.

<2> Determination of nucleotide sequence of cDNA

Clones are randomly selected from the obtained cDNA library to determine nucleotide sequences of inserts in the clones. The nucleotide sequence can be determined in accordance with the Maxam-Gilbert method or the dideoxy method. Among them, the dideoxy method is more convenient and preferred.

The nucleotide sequence can be determined in accordance with the dideoxy method by using a commercially available sequencing kit. Further, the use of an automatic sequencer makes it possible to determine sequences of a large number of clones for a short period of time.

It is unnecessary to determine the sequence for an entire length of the insert. It is enough to determine a length of nucleotide sequence which is considered to be sufficient to perform homology search. For example, in Examples described later on, the homology search as described below was performed when a sequence having not less than 60 nucleotides was successfully determined.

<3> Homology search with gene data base

The determined nucleotide sequence of each of cDNA clones is used to perform the homology search with respect to known amino acid sequences of the cellulose synthase or nucleotide sequences of genes coding therefor registered in the gene data base. The cellulose synthase is exemplified by an enzyme encoded by a gene of cellulose 4- β -glucosyltransferase (BcsA) of cellulose synthase operon of acetobacterium (Wong, H. C. et al., Proc. Natl. Acad. Sci. U.S.A., 87, 8130-8134 (1990), ACCESSION No. M37202).

Those usable as the data base include, for example, GenBank, EMBL, and DDBJ published, for example, from Los Alamos National Institute in the United States, Institute of European Molecular Biology, and National Institute of Genetics (Japan). Those commercially available and useable as the program for homology search include, for example, commercially available DNA analysis softwares, such as DNASIS (Hitachi Software Engineering Co., Ltd.) and GENE-TYX (SDC Software Development). The following methods are also available. That is, a computer terminal is connected with the host computer of National Institute of Genetics to perform analysis. Alternatively, a personal computer is connected on Internet with NCBI (National Center for Biotechnology Information) to utilize (<http://www.ncbi.nlm.nih.gov/BLAST/>) BLAST (Basic Local Alignment Search Tool) so that high speed homology search is performed.

The homology search is performed, for example, in accordance with the following algorithm. When the homology search is performed for a nucleotide sequence, homology comparison is advanced while shifting the nucleotide sequence to be investigated by every one nucleotide with respect to individual gene sequences included in the data base. When six or more continuous nucleotides are coincident, the homology score is counted and calculated in accordance with a homology score table (see, for example, M. Dayhoff, Atlas of Protein Sequence and Structure, vol. 5 (1978)). The system is set so that those having a score not less than a certain value are picked up as candidates which have homology. Further, the gap may be introduced into the sequence to be investigated or into the gene sequence included in the data base to make optimization so that the score is maximized.

When the homology search is performed for an amino acid sequence, a nucleotide sequence to be investigated is converted into amino acids concerning all six frames including those of a complementary chain. The investigation may be performed in the same manner as performed for the nucleotide. Specifically, it is possible to use blastx of BLAST described above. As for detailed techniques and conditions for the search, reference may be made to DDBJ News Letter, No. 15 (February 1995).

<4> Isolation of cDNA clone of cotton plant cellulose synthase

The clone obtained as described above is not necessarily contain the entire nucleotide sequence of the gene. In such a case, the clone is used as a probe to perform screening by means of plaque hybridization. Thus, it is possible to obtain a clone containing a full length gene from the library. A specified method may be carried out with reference to Molecular Cloning, second edition (Maniatis et al., Cold Spring Harbour Laboratory) 12.30 to 12.40.

When obtained cDNA is deficient in 5'-portion, the 5'-portion can be obtained as well by synthesizing primers so that the cDNA sequence may be elongated toward the 5'-terminal, and performing RT-PCR by using mRNA as a template.

As demonstrated in Examples described later on, the DNA of the present invention has been obtained as those having homology to the known bacterial cellulose synthase gene. The DNA further codes for an amino acid sequence GlnXXXXXXArgTrp (SEQ ID NO: 12) which is considered to form a UDP-glucose binding domain, having high homology in the vicinity thereof.

The nucleotide sequences of DNA of the present invention obtained as described above and the amino acid sequences deduced from the nucleotide sequences are shown in SEQ ID NOs: 1 to 10 in Sequence Listing. SEQ ID NOs: 1 and 3 show nucleotide sequences of PcsA1 and PcsA2 respectively. SEQ ID NOs: 2 and 4 show amino acid sequences deduced from the nucleotide sequences of PcsA1 and PcsA2 respectively.

SEQ ID NOs: 5 and 6 show a nucleotide sequence of a clone (PcsA3-682) containing 3'-side region of PcsA3 and an amino acid sequence deduced from the nucleotide sequence respectively. SEQ ID NOs: 7 and 8 show a nucleotide sequence of a 5'-portion (PcsA3-5') of another clone containing 5'-side region of PcsA3 and an amino acid sequence deduced from the nucleotide sequence respectively. SEQ ID NOs: 9 and 10 show a nucleotide sequence of 3'-portion (PcsA3-3') of the clone and an amino acid sequence deduced from the nucleotide sequence respectively (see Fig. 1). That is, SEQ ID NO: 5 corresponds to the 3'-side region of PcsA3, SEQ ID NO: 7 corresponds to the 5'-side region of PcsA3, and SEQ ID NO: 9 corresponds to internal region of PcsA3. The overlapping portion of PcsA3-682 is different from that of PcsA3-3' in 9 nucleotides in the nucleotide sequence and 1 amino acid in the amino acid sequence. Figs. 3 and 4 show the comparison between the nucleotide sequences of PcsA3-682 and PcsA3-3'. SEQ ID NO: 11 shows a combination of the amino acid sequences encoded by PcsA3-682 and PcsA3-3'.

The sequence of GlnXXXXXXArgTrp (SEQ ID NO: 12) corresponds to amino acid numbers 710 to 714 in SEQ ID NO: 2 for PcsA1, amino acid numbers 778 to 782 in SEQ ID NO: 4 for PcsA2, and amino acid numbers 356 to 360 in

SEQ ID NO: 6 for PcsA3.

PcsA1 is different from CelA1 reported by Pear et al. (Proceeding of National Academy of Science, USA (1996), 93, 12637-12642) in nucleotide sequence by 28 nucleotides. As a result, the former is different from the latter in amino acid sequence encoded thereby by 10 amino acid residues. In general, the sugar chain specificity and the substrate specificity of the sugar chain transferase are extremely changed by point mutation of the nucleotide of DNA (Yamamoto and Hakomori, The Journal of Biological Chemistry (1990) 265, 19257-19262). Therefore, it is unclear whether or not CelA1 codes for a protein having the cellulose synthase activity. Incidentally, the 48th Arg, the 56th Ser, the 81st Asn, the 104th Ala, the 110th Ser, the 247th Asp, the 376th Asp, the 386th Ser, the 409th Arg, and the 649th Ser in the amino acid sequence encoded by CelA1 correspond to Gln, Ile, Ser, Thr, Pro, Asn, Glu, Pro, His, and Gly in PcsA1 respectively.

PcsA2 of the present invention contains the same sequence as that of CelA2 reported by Pear et al. However, CelA2 has an incomplete length, and it does not contain the entire coding region. CelA2 corresponds to nucleotide numbers of 1083 to 3311 in the nucleotide sequence of PcsA2 shown in SEQ ID NO: 3.

Any of the amino acid sequences shown in SEQ ID NOs: 2, 4, 6, 8, 10, and 11 is a novel sequence. All genes having nucleotide sequences coding for the amino acid sequences are included in the present invention.

The amino acid sequences described above may include deletion, substitution, insertion, and/or addition of one or more amino acid residues provided that the characteristic of the gene of the present invention is not substantially affected. The deletion, substitution, insertion, and/or addition of one or more amino acid residues as described above is obtainable by modifying the DNA's coding for the amino acid sequences shown in SEQ ID NOs: 2, 4, 6, 8, 10, and 11 randomly in accordance with the ordinary mutation treatment or intentionally in accordance with the site-directed mutagenesis method. As described above, in general, the sugar chain specificity and the substrate specificity of the sugar chain transferase are extremely changed by point mutation of the nucleotide of DNA. Therefore, DNA coding for a protein having the cellulose synthase activity is selected from the modified DNA's. The cellulose synthase activity can be measured, for example, by means of the method described by T. Hayashi: Measuring- β -glucan deposition in plant cell walls, in Modern Methods of Plant Analysis: Plant Fibers, eds. H. F. Linskens and J. F. Jackson, Springer-Verlag, 10: 138-160 (1989).

Those harboring proteins or genes partially different from the sequences shown in Sequence Listing may exist depending on, for example, the variety of cotton plant or natural mutation. However, such genes are also included in the gene of the present invention. Such a gene may be obtained as DNA which is hybridizable under the stringent condition with all or a part of the coding region of the nucleotide sequence shown in SEQ ID NO: 1, 3, 5, 7, or 9. The "stringent condition" referred to herein indicates a condition under which a so-called specific hybrid is formed, and non-specific hybrid is not formed. It is difficult to definitely express such a condition by using a numerical value. However, for example, the stringent condition is exemplified by a condition under which nucleic acids having high homology, for example, DNA's having homology of not less than 80 % undergo hybridization with each other, and nucleic acids having homology lower than the above do not undergo hybridization with each other.

<5> Utilization of gene of the present invention

The DNA of the present invention makes it possible to control the cellulose synthesis in prokaryotic cells such as acetobacterium and/or eukaryotic cells such as yeasts belonging to, for example, the genus Saccharomyces, cells of plant such as cotton plant, and cultured cells of mammals and the like.

Specifically, the cellulose synthesis in the cells as described above can be facilitated, for example, by connecting a promoter to an upstream region of the DNA of the present invention, inserting an obtained fragment into an appropriate vector to construct a recombinant vector, and introducing the vector into the cells. Alternatively, the cellulose synthesis in the cells can be suppressed by introducing an antisense gene of the DNA of the present invention into the cells.

The promoter and the vector may be selected from those ordinarily utilized to express heterogeneous genes, and the method ordinarily employed to express heterogeneous genes may be used as the transformation method. Specifically, in the case of yeast, it is possible to use a protein-expressing kit produced by Invitrogen, i.e., Pichia Expression Kit, and a vector pPIC9 contained in this kit. For example, COS7 cells may be used as mammalian cultured cells, and a vector CDM8 may be used therefor.

The present invention provides the DNA coding for cellulose synthase. The DNA provides a new method for controlling cellulose production by incorporating the DNA into prokaryotic cells and eukaryotic cells.

Brief Description of the Drawings

Fig. 1 shows a relationship between two clones of PcsA3 as an embodiment of the DNA of the present invention. Regions interposed between arrows indicate regions for which nucleotide sequences have been determined. A dotted line indicates a region for which no nucleotide sequence has been determined.

Fig. 2 shows a structure of EcoRI adapter.

Fig. 3 shows comparison between sequences of PcsA3-682 and PcsA3-3' (former half).

Fig. 4 shows comparison between sequences of PcsA3-682 and PcsA3-3' (latter half). ":" indicates coincident nucleotides, and "*" indicates non-coincident nucleotides.

Best Mode for Carrying Out the Invention

Examples of the present invention will be explained below.

<1> Preparation of total RNA from cotton plant

Cotton plant (Gossypium hirsutum L.) Coker 312 was used as a material. Fiber cells on 16 to 18 days post anthesis were collected in liquid nitrogen. The cotton plant fiber cells in an amount of 75 g were sufficiently ground in a mortar while being frozen with liquid nitrogen. Powdered fiber was transferred to a centrifuge tube equipped with a cap, to which 375 mg of DTT as a powder was added, followed by addition of 200 ml of XT buffer (obtained by adjusting 0.2 M sodium borate containing 30 mM EDTA and 1 % SDS to be pH 9.0, and then applying a diethylpyrocarbonate treatment, followed by autoclaving to obtain a solution to which vanadylribonucleoside was added to give a concentration of 10 mM) having been heated to 90 to 95 °C. An obtained solution was sufficiently agitated.

The solution was added with 100 mg of protease K, and it was agitated again. The solution was incubated at 40 °C for 2 hours, and then it was added with 16 ml of 2 M KCl. The solution was sufficiently agitated again, and it was left to stationarily stand in ice for 1 hour, followed by centrifugation for 20 minutes (4 °C) at 12,000 g by using a high speed refrigerated centrifuge.

An obtained supernatant was filtrated, and floating matters were removed. The solution was transferred to a measuring cylinder to measure the volume. The solution was transferred to another centrifuge tube, to which lithium chloride was added in an amount of 85 mg per 1 ml of the extract solution to give a final concentration of 2 M. The solution was left to stationarily stand at 4 °C overnight, and then precipitated RNA was separated by centrifugation for 20 minutes at 12,000 g. An obtained precipitate of RNA was washed and precipitated twice with cooled 2 M lithium chloride.

The obtained RNA was dissolved in 10 mM Tris buffer (pH 7.5) to give a concentration of about 2 mg/ml, to which 5 M potassium acetate was added to give a concentration of 200 mM. Ethanol was added thereto to give a concentration of 70 %, followed by cooling at -80 °C for 10 minutes. Centrifugation was performed at 4 °C for 10 minutes at 15,000 rpm, and then an obtained precipitate was suspended in an appropriate amount of sterilized water to give an RNA sample. As a result of quantitative measurement for the RNA sample, total RNA was obtained in an amount of 2 mg.

<2> Purification of mRNA

mRNA was purified as a poly(A)⁺ RNA fraction from the total RNA obtained as described above. Purification was performed by using Oligotex-dT30 <Super> (purchased from Toyobo) as oligo(dT)-immobilized latex for poly(A)⁺ RNA purification.

Elution buffer (10 mM Tris-HCl (pH 7.5), 1 mM EDTA, 0.1 % SDS) was added to a solution containing 1 mg of the total RNA to give a total volume of 1 ml, to which 1 ml of Oligotex-dT30 <Super> was added, followed by heating at 65 °C for 5 minutes and quick cooling on ice for 3 minutes. The obtained solution was added with 0.2 ml of 5 M NaCl, and it was incubated at 37 °C for 10 minutes, followed by centrifugation at 15,000 rpm for 3 minutes. After that, a supernatant was carefully removed.

An obtained pellet was suspended in 2.5 ml of Washing Buffer (10 mM Tris-HCl (pH 7.5), 1 mM EDTA, 0.5 M NaCl, 0.1 % SDS), and the suspension was centrifuged at 15,000 rpm for 3 minutes. After that, a supernatant was carefully removed. An obtained pellet was suspended in 1 ml of TE Buffer, and then it was heated at 65 °C 5 minutes. The suspension was quickly cooled on ice for 3 minutes, and then it was centrifuged at 15,000 rpm for 3 minutes to recover poly(A)⁺ mRNA contained in an obtained supernatant.

Thus, the poly(A)⁺ mRNA in an amount of about 10 µg was obtained from 1 mg of the total RNA. An aliquot of 5 µg thereof was used to prepare a cDNA library.

<3> Preparation of cDNA library

(1) Synthesis of cDNA

The mRNA obtained as described above was used as a template to synthesis cDNA by using a λZAP cDNA synthesis kit produced by Stratagene. The following solution was prepared and mixed in a tube.

5.0 µl 10 x 1st Strand Buffer (buffer for reverse transcription reaction);
 3.0 µl 10 mM 1st Strand Methyl Nucleotide Mix (5-methyl dCTP, dATP, dGTP, dTTP mixture);
 2.0 µl Linker-Primer (linker and primer);
 H₂O (adjusted to give a total volume of 50 µl);
 1.0 µl RNase Block II (RNase inhibitor).

The respective components described above were contents of the kit. Linker-Primer had a sequence as shown in SEQ ID NO: 13. Methylated nucleotide was used because it was intended not to allow cDNA to be digested by the restriction enzyme reaction performed later on. The reaction solution was agitated well, and then 5.0 µg of poly(A)⁺ mRNA was added thereto, followed by being left to stand at room temperature for 10 minutes. Further, 2.5 µl of M-MuLV RTase (reverse transcriptase) was added (at this time, the total volume was 50 µl). The reaction solution was gently mixed, followed by centrifugation under a mild condition to allow the reaction solution to fall to the bottom of the tube. The reaction was performed at 37 °C for 60 minutes.

Next, the following solution was prepared and mixed in the tube in a certain order.

45.0 µl reaction solution containing cDNA primary chain;
 40.0 µl 10 x 2nd Strand Buffer (buffer for polymerase reaction);
 6.0 µl 2nd Strand Nucleotide Mixture (A, G, C, T mixture);
 302.0 µl H₂O.

The following solution was further added. However, in order to allow RNase and DNA polymerase to simultaneously act, enzyme solutions were allowed to adhere to the wall of the tube. After that, a vortex treatment was promptly performed, and the reaction solutions were allowed to fall to the bottom of the tube by means of centrifugation to perform a reaction for synthesizing cDNA second strand at 16 °C for 150 minutes.

0.8 µl RNase H (RNA-degrading enzyme);
 7.5 µl DNA polymerase I (10.0 u/µl).

The reaction solution was added with 400 µl of a mixed solution of phenol: chloroform (1:1). Agitation was performed well, followed by centrifugation at room temperature for 2 minutes. An obtained supernatant was added with 400 µl of phenol: chloroform again, which was subjected to a vortex treatment and centrifugation at room temperature for 2 minutes. An obtained supernatant was added with the following solution to precipitate cDNA.

33.3 µl 3 M sodium acetate solution;
 867.0 µl 100 % ethanol.

The obtained solution was left to stand at -20 °C overnight, and it was centrifuged at room temperature for 60 minutes. After that, washing was gently performed with 80 % ethanol, followed by centrifugation for 2 minutes. A supernatant was removed. An obtained pellet was dried, and it was dissolved in 43.5 µl of sterilized water. An aliquot (39.0 µl) was added with the following solution to blunt-end cDNA terminals.

5.0 µl 10 x T4 DNA Polymerase Buffer (buffer for T4 polymerase reaction);
 2.5 µl 2.5 mM dNTP Mix (A, G, C, T mixture);
 3.5 µl T4 DNA polymerase (2.9 u/µl).

The reaction was performed at 37 °C for 30 minutes, to which 50 µl of distilled water was added, and then 100 µl of phenol: chloroform was added thereto, followed by a vortex treatment and centrifugation for 2 minutes. An obtained supernatant was added with 100 µl of chloroform, which was subjected to a vortex treatment, followed by centrifugation for 2 minutes. The supernatant was added with the following solution to precipitate cDNA.

7.0 µl 3 M sodium acetate solution;
 226 µl 100 % ethanol.

The solution was left to stand on ice for 30 minutes or more, and it was centrifuged at 4 °C for 60 minutes. An obtained precipitate was washed with 150 µl of 80 % ethanol, followed by centrifugation for 2 minutes and drying. The cDNA pellet was dissolved in 7.0 µl of EcoRI Adaptor solution, to which the following solution was added to ligate the EcoRI adaptor to both ends of the cDNA. Sequences of respective strands of the EcoRI adaptor are shown in SEQ ID NO: 14 and Fig. 2.

1.0 µl 10 x Ligation Buffer (buffer for ligase reaction);
 1.0 µl 10 mM ATP;
 1.0 µl T4 DNA ligase.

5 The reaction solution was centrifuged under a mild condition, and it was left to stand at 4 °C overnight or more. The solution was treated at 70 °C for 30 minutes, and then it was centrifuged under a mild condition, followed by being left to stand at room temperature for 5 minutes. The reaction solution was added with the following solution to phosphorylate 5'-terminals of the EcoRI adapter.

10 1.0 µl 10 x Ligation Buffer (buffer for ligase reaction);
 2.0 µl 10 mM ATP;
 6.0 µl H₂O;
 1.0 µl T4 polynucleotide kinase (10.0 u/µl).

15 The reaction was performed at 37 °C for 30 minutes, followed by a treatment at 70 °C for 30 minutes. The solution was centrifuged under a mild condition, and it was left to stand at room temperature for 5 minutes. The following solution was further added thereto to perform a reaction at 37 °C for 90 minutes so that the XhoI site introduced by Linker-Primer was digested with XhoI, followed by being left to stand at room temperature to perform cooling.

20 28.0 µl XhoI Buffer;
 3.0 µl XhoI (45 u/µl).

The reaction solution was added with 5.0 µl of 10 x STE (10 mM Tris-HCl (pH 8.0), 100 mM NaCl, 1 mM EDTA), which was added into a centrifuge column for removing short fragments (Sephacryl Spin Column) to perform centrifugation at 600 g for 2 minutes to obtain an eluent which was designated as Fraction 1. This operation was further repeated three times to obtain Fractions 2, 3, and 4 respectively. Fractions 3 and 4 were combined, to which phenol:chloroform (1:1) was added and agitated well, followed by centrifugation at room temperature for 2 minutes. An obtained supernatant was added with an equal amount of chloroform, and an obtained mixture was agitated well. The mixture was centrifuged at room temperature for 2 minutes to obtain a supernatant to which a two-fold amount of 100 % ethanol was added, followed by being left to stand at - 20 °C overnight. The solution was centrifuged at 4 °C for 60 minutes, followed by washing with an equal amount of 80 % ethanol. The solution was centrifuged at 4 °C for 60 minutes to obtain a cDNA pellet which was suspended in 10 µl of sterilized water.

(2) Preparation of cDNA library

35 The double strand cDNA obtained as described above was ligated with λ phage expression vector to prepare a recombinant vector. The following solution was prepared and mixed in a tube to perform a reaction at 12 °C overnight, followed by being left to stand at room temperature for 2 hours to ligate cDNA with the vector.

40 2.5 µl cDNA solution;
 0.5 µl 10 x Ligation Buffer;
 0.5 µl 10 mM ATP;
 1.0 µl λZAP vector DNA (1 µg/µl);
 0.5 µl T4 DNA ligase (4 Weiss u/µl).

(3) Packaging of phage DNA into phage particles

50 The phage vector containing the cDNA was packaged into phage particles by using an in vitro packaging kit (Gigapack II Gold packaging extract: produced by Stratagene). The recombinant phage solution was added to Freeze/Thaw extract immediately after dissolution, and the solution was placed on ice, to which 15 µl of Sonic extract was added to perform mixing well by pipetting. The reaction solution was centrifuged under a mild condition, and it was left to stand at room temperature (22 °C) for 2 hours. The reaction solution was added with 500 µl of Phage Dilution Buffer, to which 20 µl of chloroform was further added, followed by mixing. In order to measure the titer of the library, an aliquot (2 µl) of 500 µl of the aqueous phase was diluted in a ratio of 1:10 with 18 µl of SM buffer (5.8 g of NaCl, 2 g of MgSO₄•7H₂O, 50 ml of 1 M Tris-HCl (pH 7.5), and 5 ml of 2 % gelatin in 1 L). The diluted solution (1 µl) and the phage stock solution (1 µl) were plated respectively together with 200 p1 of a culture solution of Escherichia coli PLK-F' strain having been cultivated to arrive at a value of OD₆₀₀ of 0.5. That is, Escherichia coli PLK-F' strain was mixed with the phage solution to perform cultivation at 37 °C for 15 minutes. The obtained culture was added to 2 to 3 ml of top agar

(48 °C), which was immediately overlaid on NZY agar plate having been warmed at 37 °C. Cultivation was performed overnight at 37 °C, and appeared plaques were counted to calculate the titer. As a result, the titer was 1.2×10^6 pfu/ml.

(4) Amplification of library

A centrifuge tube was added with the packaging solution containing about 50,000 recombinant bacteriophages and 600 µl of a culture solution of *Escherichia coli* PLK-F' strain having been cultivated to have a value of OD₆₀₀ of 0.5, followed by cultivation at 37 °C for 15 minutes. The culture solution was added with 6.5 ml of top agar having been maintained at 48 °C after dissolution, which was overlaid on 150 mm NZY plate having been warmed at about 37 °C, followed by cultivation at 37 °C for 5 to 8 hours. The respective plates were added with 10 ml of SM Buffer to perform cultivation at 4 °C overnight with gentle shaking. SM Buffer in the respective plates was collected in a sterilized polypropylene tube. The respective plates were rinsed with 2 ml of SM Buffer, and the rinsing solutions were collected in the same tube. Chloroform in an amount corresponding to 5 % of the total amount was added and mixed, followed by being left to stand at room temperature for 15 minutes. Bacterial cells were removed by centrifugation at 4,000 g for 5 minutes. An obtained supernatant was added with chloroform in an amount corresponding to 0.3 % of the total amount, and it was stored at 4 °C. The titer of the library amplified as described above was measured in the same manner as described above. As a result, the titer was 2.3×10^9 pfu/ml.

(5) Excision of plasmid from phage DNA

In vivo excision of the plasmid portion from the recombinant phage DNA was performed. The following solution was mixed in 50 ml of a conical tube to cause infection at 37 °C for 15 minutes:

culture solution of *Escherichia coli* XL1-Blue (OD₆₀₀ = 0.1) 200 µl;
phage solution after amplification 200 µl ($> 1 \times 10^5$ phage particles);
helper phage R408 1 µl ($> 1 \times 10^6$ pfu/ml).

The mixed solution was added with 5 ml of 2 x YT medium to perform cultivation at 37 °C for 3 hours with shaking. A heat treatment was applied thereto at 70 °C for 20 minutes, followed by centrifugation at 4,000 g for 5 minutes. An obtained supernatant was decanted and transferred to a sterilized tube. Centrifugation was performed to obtain a supernatant which was diluted 100 times to obtain a solution. An aliquot (20 µl) of the solution was mixed with 200 µl of a culture solution of *Escherichia coli* XL1-Blue having been cultivated to obtain a value of OD₆₀₀ of 1.0 to cause infection at 37 °C for 15 minutes. Aliquots (1 to 100 µl) of the culture solution were plated on LB plates containing ampicillin, followed by cultivation at 37 °C overnight. Appeared colonies were randomly selected. Selected colonies were added with glycerol, and they were stored at -80 °C.

(6) Preparation of plasmid

Plasmids were prepared by using Magic Mini-prep kit produced by Promega. The culture fluid of *Escherichia coli* harboring the plasmid having been stored at -80 °C was inoculated into 5 ml of 2 x YT medium, followed by cultivation at 37 °C overnight. Centrifugation was performed for 5 minutes (4,000 rpm, 4 °C), and a supernatant was removed by decantation. An obtained bacterial cell pellet was added with 1 ml of TE buffer, followed by a vortex treatment. An obtained bacterial cell suspension was transferred to an Eppendorf tube, followed by centrifugation for 5 minutes (5,000 rpm, 4 °C). A resultant supernatant was removed by decantation.

An obtained bacterial cell pellet was added with 300 µl of Cell Resuspension Solution, and it was sufficiently suspended therein. An obtained suspension was transferred to an Eppendorf tube. The suspension was agitated for 2 minutes with a mixer, to which 300 µl of Cell Lysis Solution was added, followed by agitation until the suspension became transparent. Neutralization Solution (300 µl) was added thereto, and agitation was performed by shaking with the hand, followed by centrifugation for 10 minutes (15,000 rpm).

Only an obtained supernatant was transferred to a new Eppendorf tube (1.5 ml). A suction tube was prepared, to which a cock, a miniature column and a syringe (injector) were connected in this order. A resin in an amount of 1 ml was charged into the syringe. The supernatant was poured into the syringe, and agitation was performed well, followed by suction. Column Washing Solution in an amount of 2 ml was added, and washing was performed while performing suction. Suction was continued for 1 to 2 minutes in order to dry up. The miniature column was removed from the equipment, and it was set in a new Eppendorf tube (1.5 ml). Sterilized water in an amount of 100 µl having been warmed at 65 to 70 °C was poured into the miniature column, and the column and the Eppendorf tube were centrifuged together for 1 minute (5,000 rpm). An eluted solution was transferred to an Eppendorf tube, to which 5 µl of 3 M sodium acetate aqueous solution was added, and 250 µl of cold ethanol was added thereto. The solution was centrifuged (15,000 rpm,

25 minutes), and a supernatant was discarded. An obtained precipitate was added with 1 ml of 70 % ethanol, followed by centrifugation again (15,000 rpm, 3 minutes). Ethanol was completely removed, and the tube was vacuum-dried in a desiccator. The precipitate was sufficiently dissolved in 20 µl of sterilized water, and an obtained solution was stored at -20 °C. An aliquot (1 µl) of the solution was dispensed, and it was subjected to electrophoresis together with volume markers to quantitatively determine the plasmid DNA.

<4> Determination of nucleotide sequence of cDNA and homology search with gene data base

(1) Determination of nucleotide sequence of cDNA

The nucleotide sequence of cDNA was analyzed by using DNA automatic sequencer 373A produced by Applied Biosystems Inc. (ABI). The sequencing reaction was performed in accordance with an attached manual by using T3 primer based on the use of Dye Primer Cycle Sequencing Kit produced by the same company. The nucleotide sequence was determined for about 750 clones which were randomly selected.

(2) Homology search

Partial sequences of about 750 clones were searched with a computer using BlastX. As a result, three clones appeared to be homologues of bacterial cellulose synthase subunit. Therefore, it was tried to isolate full length clones.

<5> Isolation of full length clones

(1) 5'-RACE

As a result of the homology search, the obtained homologue clones were found to be partial length clones. Therefore, primers were synthesized to make elongation toward the 5' upstream so that RT-PCR was performed by using mRNA as a template.

(1-a) Synthesis of first-strand DNA

The following solution was prepared and mixed in a tube.

0.5 µl 10 µmol gene-specific primer 1;
1 pg total RNA;
DEPC-treated H₂O (adjusted to give a total amount of 9 µl).

The following oligonucleotides were used as the gene-specific primer 1. That is, an oligonucleotide having a nucleotide sequence shown in SEQ ID NO: 15 was used for PcsA1. An oligonucleotide having a nucleotide sequence shown in SEQ ID NO: 16 was used for PcsA2. An oligonucleotide having a nucleotide sequence shown in SEQ ID NO: 17 was used for PcsA3.

The reaction solution was gently mixed, and then it was centrifuged under a mild condition to allow the reaction solution to fall to the bottom of the tube. The solution was left to stand at 70 °C for 10 minutes, followed by immediate cooling on ice.

Next, the following solution was prepared and mixed in the tube.

5 x RT Buffer 5 p1;
25 mM MgCl₂ 2.5 µl;
2 mM dNTP mix 5 µl;
0.1 M DTT 2.5 µl;
H₂O (added to give a total amount of 24 µl).

The solution was gently agitated, and then it was centrifuged under a mild condition to allow the reaction solution to fall to the bottom of the tube, followed by being left to stand at 42 °C for 1 minute. The solution was added with 1 µl of SuperScriptII RT (reverse transcriptase, GIBCO BRL), and it was gently mixed. After that, the reaction was performed at 42 °C for 50 minutes. Subsequently, the reaction solution was left to stand at 70 °C for 15 minutes to stop the reaction. Centrifugation was performed under a mild condition to allow the reaction solution to fall to the bottom of the tube, followed by being left to stand at 37 °C. RNase H (produced by Toyobo) in an amount of 1 µl was added thereto to perform a reaction at 37 °C for 30 minutes.

Subsequently, in order to remove excessive primers and nucleotides contained in the reaction solution, gel filtration was performed by using a purification column produced by Boehringer, Quick Spin Columns. At first, the tip of the column was removed, followed by centrifugation at 1,100 x g for 2 minutes to discard the buffer. The reaction solution was introduced into the central area of the column, followed by centrifugation at 1,100 x g for 4 minutes to recover the solution.

(1-b) Poly(dC) tailing

An aliquot (5 µl) was dispensed from the obtained solution, to which the following solution was added.

5 µl 5 x CoCl₂ Buffer;
2.5 µl 2 mM dCTP;
H₂O (adjusted to give a total amount of 24 µl).

The reaction solution was mixed well, and it was left to stand at 94 °C for 3 minutes. Centrifugation was performed under a mild condition to allow the reaction solution to fall to the bottom of the tube, followed by being left to stand on ice. Terminal transferase TdT (produced by Toyobo) was added thereto in an amount of 1 µl, followed by mixing under a mild condition to perform a reaction at 37 °C for 10 minutes. Subsequently, the reaction solution was left to stand at 65 °C for 10 minutes to stop the reaction.

(1-c) PCR reaction

An aliquot (2.5 µl) was dispensed from the reaction solution, to which the following solution was added.

2.5 µl 10 x PCR Buffer;
2.5 µl 2 mM dNTP mix;
0.5 µl Gene-specific primer 2;
0.5 µl Abridged Anchor Primer (GIBCO BRL);
0.5 µl Advantage KlenTaq Polymerase Mix (Clontech);
H₂O (adjusted to give a total amount of 25 µl).

The following oligonucleotides were used as Gene-specific primer 2. That is, an oligonucleotide having a nucleotide sequence shown in SEQ ID NO: 18 was used for PcsA1. An oligonucleotide having a nucleotide sequence shown in SEQ ID NO: 19 was used for PcsA2. An oligonucleotide having a nucleotide sequence shown in SEQ ID NO: 20 was used for PcsA3.

The solution was introduced into a 0.2 ml tube to perform the PCR reaction under the following condition.

PAD	94 °C	90 seconds
30 cycles	94 °C	30 seconds
	60 to 68 °C	30 to 60 seconds
	68 °C	180 seconds
Final	68 °C	7 minutes
Hold	4 °C	

The reaction solution was subjected to agarose gel electrophoresis to extract, from the gel, DNA's corresponding to portions having the largest size (about 1.8 K for PcsA1, about 2 K for PcsA2, and about 2.2 K for PcsA3). GENO-BIND produced by CLONTECH was used for the extraction, and the procedure was carried out in accordance with its protocol. The DNA thus obtained was subjected to Poly(dC) tailing, which was used as a template to perform the PCR reaction. The condition and the composition of the reaction solution were the same as those described above.

(2) Cloning

(2-a) 5'-RACE TA cloning

Starting from the obtained PCR reaction solution, cloning was performed by using TA Cloning Kit produced by Invitrogen in accordance with its protocol.

The following solution was added to an aliquot (1.5 µl) of the PCR reaction solution obtained as described above.

0.5 µl 10 x Ligation Buffer;
 1 µl pCRII vector;
 0.5 µl T4 DNA Ligase;
 1.5 µl dH₂O.

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The reaction was performed at 14 °C overnight. An aliquot (2 p1) of the reaction solution was added to 25 p1 of *Escherichia coli* competent cell (JM109) preparation, followed by being left to stand for 30 minutes on ice. After that, heat shock was applied at 42 °C for 30 seconds. The solution was stationarily left to stand on ice for 2 minutes, to which 450 µl of SOB medium was thereafter added to perform cultivation at 37 °C for 1 hour with shaking at 200 rpm.

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The culture was spread over Amp/Xgal/IPTG plate, followed by incubation at 37 °C overnight. The plasmid was extracted from obtained colonies in accordance with the method as described above.

(2-b) Cloning of complete length cDNA

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The procedure was carried out by using DNA Sequencer 377 produced by ABI in accordance with its protocol. The sequencing reaction was performed by using M13 primer and synthetic oligomer as primers, based on the use of Dye Terminator Cycle Sequencing Kit produced by the same company. As a result of the sequencing, as for PcsA3, it was revealed that another clone also belonging to the group of PcsA3 but having a slightly different sequence (one position for amino acid) was isolated (see Figs. 3 and 4). A nucleotide sequence of a clone (PcsA3-682) containing the 3'-side region of PcsA3 and an amino acid sequence deduced from this nucleotide sequence are shown in SEQ ID NOs: 5 and 6. A nucleotide sequence of a 5'-portion (PcsA3-5') of another clone containing the 5'-side region of PcsA3 and an amino acid sequence deduced from this nucleotide sequence are shown in SEQ ID NOs: 7 and 8. A nucleotide sequence of a 3'-portion (PcsA3-3') of the clone and an amino acid sequence deduced from this nucleotide sequence are shown in SEQ ID NOs: 9 and 10.

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As for PcsA1 and PcsA2, primers for 5'-terminal and 3'-terminal of a region containing ORF were synthesized on the basis of the obtained sequences to perform the PCR reaction. Thus, complete length clones were isolated by means of TA cloning. The condition and the composition of the reaction solution were the same as those described above.

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Oligonucleotides shown in SEQ ID NO: 21 (5'-terminal) and SEQ ID NO: 22 (3'-terminal) were used as the primers for PcsA1. Oligonucleotides shown in SEQ ID NO: 23 (5'-terminal) and SEQ ID NO: 24 (3'-terminal) were used as the primers for PcsA2. Results are shown in SEQ ID NOs: 1 to 4.

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Ann x to th description

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: NISSHINBO INDUSTRIES, INC.

HAYASHI, Takahisa

(ii) TITLE OF INVENTION: CELLULOSE SYNTHASE GENE

(iii) NUMBER OF SEQUENCES: 24

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE:

(B) STREET:

(C) CITY:

(E) COUNTRY:

(F) ZIP:

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk

(B) COMPUTER: IBM PC compatible

(C) OPERATING SYSTEM: PC-DOS/MS-DOS

(D) SOFTWARE: PatentIn Release #1.0, Version #1.30

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:

(B) FILING DATE:

(C) CLASSIFICATION:

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: JP 9-83133

(B) FILING DATE: 1-APR-1997

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME:

(B) REGISTRATION NUMBER:

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE:

(B) TELEFAX:

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3207 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Gossypium hirsutum L.

(C) INDIVIDUAL ISOLATE: Coker312

(1x) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 77..3001

(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

	GGTTAGCATA TTGTTTGTAG CATTGGGTTT TTTTCTCAAG GAAGAAGAAG GAGAAAGATA	60
	AGTAATGTTT TTGAGA ATG ATG GAA TCT GGG GTT OCT GTT TGC CAC ACT	109
10	Met Met Glu Ser Gly Val Pro Val Cys His Thr	
	1 5 10	
	TGT GGT GAA CAT GTT GGG TTG AAT GTT AAT GGT GAA CCC TTT GTG GCT	157
	Cys Gly Glu His Val Gly Leu Asn Val Asn Gly Glu Pro Phe Val Ala	
15	15 20 25	
	TGC CAT GAA TGT AAT TTC OCT ATT TGT AAG AGT TGT TTT GAG TAT GAT	205
	Cys His Glu Cys Asn Phe Pro Ile Cys Lys Ser Cys Phe Glu Tyr Asp	
	30 35 40	
20	CTT AAG GAA GGA CAA AAA GCT TGC TTG CGT TGT GGT ATT CCG TAT GAT	253
	Leu Lys Glu Gly Gln Lys Ala Cys Leu Arg Cys Gly Ile Pro Tyr Asp	
	45 50 55	
	GAA AAC CTG TTG GAC GAT GTC GAG AAG GOC ACC GGC GAT CAA TCG ACA	301
25	Glu Asn Leu Leu Asp Asp Val Glu Lys Ala Thr Gly Asp Gln Ser Thr	
	60 65 70 75	
	ATG GCT GCA CAT TTG AGC AAG TCT CAG GAT GTT GGA ATT CAT GCA AGA	349
	Met Ala Ala His Leu Ser Lys Ser Gln Asp Val Gly Ile His Ala Arg	
30	80 85 90	
	CAT ATC AGC AGT GTG TCT ACA TTG GAT AGT GAA ATG ACT GAA GAC AAT	397
	His Ile Ser Ser Val Ser Thr Leu Asp Ser Glu Met Thr Glu Asp Asn	
	95 100 105	
35	GGG AAT CCG ATT TGG AAG AAC AGG GTG GAA AGT TGG AAA GAA AAG AAG	445
	Gly Asn Pro Ile Trp Lys Asn Arg Val Glu Ser Trp Lys Glu Lys Lys	
	110 115 120	
	AAC AAG AAG AAG AAG OCT GCA ACA ACT AAG GTT GAA AGA GAG GCT GAA	493
40	Asn Lys Lys Lys Lys Pro Ala Thr Thr Lys Val Glu Arg Glu Ala Glu	
	125 130 135	
	ATC OCA OCT GAG CAA CAA ATG GAA GAT AAA CCG GCA CCG GAT GCT TOC	541
	Ile Pro Pro Glu Gln Gln Met Glu Asp Lys Pro Ala Pro Asp Ala Ser	
45	140 145 150 155	
	CAG CCC CTC TCG ACT ATA ATT CCA ATC CCG AAA AGC AGA CTT GCA CCA	589
	Gln Pro Leu Ser Thr Ile Ile Pro Ile Pro Lys Ser Arg Leu Ala Pro	
	160 165 170	
50	TAC OGA ACC GTG ATC ATT ATG CGA TTG ATC ATT CTC GGT CTT TTC TTC	637
	Tyr Arg Thr Val Ile Ile Met Arg Leu Ile Ile Leu Gly Leu Phe Phe	

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		175		180		185											
	CAT	TAT	OGA	GTA	ACA	AAC	CCC	GTT	GAC	AGT	GCT	TTT	GGA	CTG	TGG	CTC	685
5	His	Tyr	Arg	Val	Thr	Asn	Pro	Val	Asp	Ser	Ala	Phe	Gly	Leu	Trp	Leu	
			190					195					200				
	ACT	TCA	GTC	ATA	TGT	GAA	ATC	TGG	TTT	GCT	TTT	TOC	TGG	GTG	TTG	GAT	733
	Thr	Ser	Val	Ile	Cys	Glu	Ile	Trp	Phe	Ala	Phe	Ser	Trp	Val	Leu	Asp	
			205					210					215				
10	CAG	TTC	OCT	AAG	TGG	TAT	OCT	GTT	AAC	AGG	GAA	ACA	TAC	ATT	GAC	AGA	781
	Gln	Phe	Pro	Lys	Trp	Tyr	Pro	Val	Asn	Arg	Glu	Thr	Tyr	Ile	Asp	Arg	
	220					225					230				235		
	CTG	TCT	GCA	AGA	TAT	GAA	AGA	GAA	GGT	GAA	OCT	AAT	GAA	CTT	GCT	GCA	829
15	Leu	Ser	Ala	Arg	Tyr	Glu	Arg	Glu	Gly	Glu	Pro	Asn	Glu	Leu	Ala	Ala	
				240					245					250			
	GTT	GAC	TTC	TTT	GTG	AGT	ACA	GTG	GAT	OCA	TTG	AAA	GAG	OCT	OCA	TTG	877
	Val	Asp	Phe	Phe	Val	Ser	Thr	Val	Asp	Pro	Leu	Lys	Glu	Pro	Pro	Leu	
20				255					260					265			
	ATT	ACT	GCC	AAT	ACT	GTG	CTT	TOC	ATC	CTT	GCC	TTG	GAC	TAC	COG	GTA	925
	Ile	Thr	Ala	Asn	Thr	Val	Leu	Ser	Ile	Leu	Ala	Leu	Asp	Tyr	Pro	Val	
				270					275					280			
25	GAT	AAG	GTC	TCT	TGT	TAT	ATA	TCT	GAT	GAT	GGT	GCG	GCC	ATG	CTG	ACA	973
	Asp	Lys	Val	Ser	Cys	Tyr	Ile	Ser	Asp	Asp	Gly	Ala	Ala	Met	Leu	Thr	
				285					290					295			
	TTT	GAA	TCT	CTA	GTA	GAA	ACA	GCC	GAC	TTT	GCA	AGA	AAG	TGG	GTT	CCA	1021
30	Phe	Glu	Ser	Leu	Val	Glu	Thr	Ala	Asp	Phe	Ala	Arg	Lys	Trp	Val	Pro	
	300					305					310				315		
	TTC	TGC	AAA	AAA	TTT	TCC	ATT	GAA	OCA	CGG	GCA	OCT	GAG	TTT	TAC	TTC	1069
	Phe	Cys	Lys	Lys	Phe	Ser	Ile	Glu	Pro	Arg	Ala	Pro	Glu	Phe	Tyr	Phe	
35				320						325				330			
	TCA	CAG	AAG	ATT	GAT	TAC	TTG	AAA	GAT	AAA	GTG	CAG	CCC	TCT	TTT	GTA	1117
	Ser	Gln	Lys	Ile	Asp	Tyr	Leu	Lys	Asp	Lys	Val	Gln	Pro	Ser	Phe	Val	
				335					340					345			
40	AAA	GAA	CGT	AGA	GCT	ATG	AAA	AGA	GAT	TAC	GAA	GAG	TAC	AAA	ATT	CGA	1165
	Lys	Glu	Arg	Arg	Ala	Met	Lys	Arg	Asp	Tyr	Glu	Glu	Tyr	Lys	Ile	Arg	
				350					355					360			
	ATC	AAT	GCT	TTA	GTT	GCA	AAG	GCT	CAG	AAA	ACA	OCT	GAA	GAA	GGA	TGG	1213
45	Ile	Asn	Ala	Leu	Val	Ala	Lys	Ala	Gln	Lys	Thr	Pro	Glu	Glu	Gly	Trp	
				365					370					375			
	ACA	ATG	CAA	GAT	GGA	ACT	OCT	TGG	COG	GGA	AAT	AAC	COG	CGT	GAT	CAC	1261
	Thr	Met	Gln	Asp	Gly	Thr	Pro	Trp	Pro	Gly	Asn	Asn	Pro	Arg	Asp	His	
50	380					385					390				395		
	OCT	GGC	ATG	ATT	CAG	GTT	TTC	CTT	GGA	TAT	AGC	GGT	GCT	CAT	GAC	ATC	1309

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	Pro Gly Met Ile Gln Val Phe Leu Gly Tyr Ser Gly Ala His Asp Ile	
	400 405 410	
5	GAA GGA AAT GAA CTT OCC OGA CTG GTT TAC GTC TCT AGA GAG AAG AGA	1357
	Glu Gly Asn Glu Leu Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg	
	415 420 425	
10	OCT GGC TAC CAA CAC CAC AAA AAG GCT GGT GCT GAA AAT GCT TTG GTT	1405
	Pro Gly Tyr Gln His His Lys Lys Ala Gly Ala Glu Asn Ala Leu Val	
	430 435 440	
	AGG GTG TCT GCA GTT CTT ACA AAT GCT OCC TTC ATC CTC AAT CTT GAT	1453
	Arg Val Ser Ala Val Leu Thr Asn Ala Pro Phe Ile Leu Asn Leu Asp	
	445 450 455	
15	TGT GAC CAC TAT GTT AAC AAT AGC AAG GCA GTT AGG GAG GCA ATG TGC	1501
	Cys Asp His Tyr Val Asn Asn Ser Lys Ala Val Arg Glu Ala Met Cys	
	460 465 470 475	
20	TTC TTG ATG GAC CCA CAA GTC GGT OGA GAT GTC TGC TAT GTG CAG TTT	1549
	Phe Leu Met Asp Pro Gln Val Gly Arg Asp Val Cys Tyr Val Gln Phe	
	480 485 490	
	OCT CAA AGA TTT GAT GGC ATA GAT AGG AGT GAT OGA TAT GCC AAT CGG	1597
	Pro Gln Arg Phe Asp Gly Ile Asp Arg Ser Asp Arg Tyr Ala Asn Arg	
25	495 500 505	
	AAC ACA GTT TTC TTT GAT GTT AAC ATG AAA GGT CTT GAT GGA ATC CAA	1645
	Asn Thr Val Phe Phe Asp Val Asn Met Lys Gly Leu Asp Gly Ile Gln	
	510 515 520	
30	GGG OCT GTT TAT GTG GGA ACA GGT TGT GTT TTC AAT AGG CAA GCA CTT	1693
	Gly Pro Val Tyr Val Gly Thr Gly Cys Val Phe Asn Arg Gln Ala Leu	
	525 530 535	
35	TAT GGC TAT GGT OCA OCT TCA ATG OCA AGT TTT OCC AAG TCA TOC TOC	1741
	Tyr Gly Tyr Gly Pro Pro Ser Met Pro Ser Phe Pro Lys Ser Ser Ser	
	540 545 550 555	
	TCA TCT TGC TCG TGT TGC TGC OCC GGC AAG AAG GAA OCT AAA GAT CCA	1789
	Ser Ser Cys Ser Cys Cys Cys Pro Gly Lys Lys Glu Pro Lys Asp Pro	
40	560 565 570	
	TCA GAG CTT TAT AGG GAT GCA AAA CGG GAA GAA CTT GAT GCT GCC ATC	1837
	Ser Glu Leu Tyr Arg Asp Ala Lys Arg Glu Glu Leu Asp Ala Ala Ile	
	575 580 585	
45	TTT AAC CTT AGG GAA ATT GAC AAT TAT GAT GAG TAT GAA AGA TCA ATG	1885
	Phe Asn Leu Arg Glu Ile Asp Asn Tyr Asp Glu Tyr Glu Arg Ser Met	
	590 595 600	
50	TTG ATC TCT CAA ACA AGC TTT GAG AAA ACT TTT GGC TTA TCT TCA GTC	1933
	Leu Ile Ser Gln Thr Ser Phe Glu Lys Thr Phe Gly Leu Ser Ser Val	
	605 610 615	

	TTC ATT GAA TCT ACA CTA ATG GAG AAT GGA GGA GTG GCT GAA TCT GGC	1981
	Phe Ile Glu Ser Thr Leu Met Glu Asn Gly Gly Val Ala Glu Ser Ala	
5	620 625 630 635	
	AAC CCT TOC ACA CTA ATC AAG GAA GCA ATT CAT GTC ATC GGC TGT GGC	2029
	Asn Pro Ser Thr Leu Ile Lys Glu Ala Ile His Val Ile Gly Cys Gly	
	640 645 650	
10	TAT GAG GAG AAG ACT GCA TGG GGG AAA GAG ATT GGA TGG ATA TAT GGT	2077
	Tyr Glu Glu Lys Thr Ala Trp Gly Lys Glu Ile Gly Trp Ile Tyr Gly	
	655 660 665	
	TCA GTC ACT GAG GAT ATC TTA AOC GGC TTC AAA ATG CAC TGC CGA GGA	2125
15	Ser Val Thr Glu Asp Ile Leu Thr Gly Phe Lys Met His Cys Arg Gly	
	670 675 680	
	TGG AGA TOG ATT TAC TGC ATG OOC TTA AGG CCA GCA TTC AAA GGA TCT	2173
	Trp Arg Ser Ile Tyr Cys Met Pro Leu Arg Pro Ala Phe Lys Gly Ser	
	685 690 695	
20	GCA OOC ATC AAT CTG TCT GAT OGG TTG CAC CAG GTT CTT CGA TGG GCT	2221
	Ala Pro Ile Asn Leu Ser Asp Arg Leu His Gln Val Leu Arg Trp Ala	
	700 705 710 715	
	CTT GGA TCT GTT GAA ATT TTC CTA AGC AGG CAT TGC OCT CTA TGG TAT	2269
25	Leu Gly Ser Val Glu Ile Phe Leu Ser Arg His Cys Pro Leu Trp Tyr	
	720 725 730	
	GGC TTT GGA GGT GGT OGT CTT AAA TGG CTT CAA AGA CTA GCA TAT ATA	2317
	Gly Phe Gly Gly Gly Arg Leu Lys Trp Leu Gln Arg Leu Ala Tyr Ile	
30	735 740 745	
	AAC AOC ATT GTC TAT OCT TTC ACA TOC CTT OCA CTC ATT GOC TAT TGT	2365
	Asn Thr Ile Val Tyr Pro Phe Thr Ser Leu Pro Leu Ile Ala Tyr Cys	
	750 755 760	
35	TCA CTA CCA GCA ATC TGT CTT CTC ACA GGA AAA TTT ATC ATA CCA ACG	2413
	Ser Leu Pro Ala Ile Cys Leu Leu Thr Gly Lys Phe Ile Ile Pro Thr	
	765 770 775	
	CTC TCA AAC CTG GCA AGT GTT CTC TTT CTT GGC CTT TTC CTT TOC ATT	2461
40	Leu Ser Asn Leu Ala Ser Val Leu Phe Leu Gly Leu Phe Leu Ser Ile	
	780 785 790 795	
	ATC GTG ACT GCT GTT CTC GAG CTC CGA TGG AGT GGT GTC AGC ATT GAG	2509
	Ile Val Thr Ala Val Leu Glu Leu Arg Trp Ser Gly Val Ser Ile Glu	
45	800 805 810	
	GAC TTA TGG OGT AAC GAG CAG TTT TGG GTC ATC GGT GGC GTT TCA GOC	2557
	Asp Leu Trp Arg Asn Glu Gln Phe Trp Val Ile Gly Gly Val Ser Ala	
	815 820 825	
50	CAT CTC TTT GOC GTC TTC CAA GGT TTC CTT AAG ATG CTT GCG GGC ATT	2605
	His Leu Phe Ala Val Phe Gln Gly Phe Leu Lys Met Leu Ala Gly Ile	

55

	830	835	840	
	GAC ACC AAC TTT ACT GTC ACT GGC AAA GCA GCT GAT GAT GCA GAT TTT			2653
5	Asp Thr Asn Phe Thr Val Thr Ala Lys Ala Ala Asp Asp Ala Asp Phe			
	845	850	855	
	GGT GAG CTC TAC ATT GTG AAA TGG ACT ACA CTT CTA ATC OCT OCA ACA			2701
	Gly Glu Leu Tyr Ile Val Lys Trp Thr Thr Leu Leu Ile Pro Pro Thr			
10	860	865	870	875
	ACA CTC CTC ATC GTC AAC ATG GTT GGT GTC GTT GGC GGA TTC TOC GAT			2749
	Thr Leu Leu Ile Val Asn Met Val Gly Val Val Ala Gly Phe Ser Asp			
	880	885	890	
15	GGC CTC AAC AAA GGG TAC GAA GCT TGG GGA OCA CTC TTT GGC AAA GTG			2797
	Ala Leu Asn Lys Gly Tyr Glu Ala Trp Gly Pro Leu Phe Gly Lys Val			
	895	900	905	
	TTC TTT TOC TTC TGG GTC ATC CTC CAT CTT TAT CCA TTC CTC AAA GGT			2845
	Phe Phe Ser Phe Trp Val Ile Leu His Leu Tyr Pro Phe Leu Lys Gly			
20	910	915	920	
	CTT ATG GGA CGC CAA AAC AGG ACA OCA ACC ATT GTT GTC CTT TGG TCA			2893
	Leu Met Gly Arg Gln Asn Arg Thr Pro Thr Ile Val Val Leu Trp Ser			
	925	930	935	
25	GTG TTG TTG GCT TCT GTC TTC TCT CTT GTT TGG GTT CGG ATC AAC CCG			2941
	Val Leu Leu Ala Ser Val Phe Ser Leu Val Trp Val Arg Ile Asn Pro			
	940	945	950	955
	TTT GTC AGC ACC GGC GAT AGC ACC ACC GTG TCA CAG AGC TGC ATT TOC			2989
30	Phe Val Ser Thr Ala Asp Ser Thr Thr Val Ser Gln Ser Cys Ile Ser			
	960	965	970	
	ATT GAT TGT TGATGATATT ATGTGTTTCT TAGAATTGAA ATCATTGCAA			3038
	Ile Asp Cys			
35	GTAAGTGGAC TGAAACATGT CTATTGACTA AGTTTTGAAC AGTTTGTAOC CATTTTATTC			3098
	TTAGCAGTGT GTAATTTTTC TAAACAATGC TATGAACTAT ACATATTTCA TTGATATTTA			3158
	CATTAAATGA AACTACATCA GTCTGCAGAA AAAAAAAAAA AAAAAAAAAA			3207

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 974 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Met	Met	Glu	Ser	Gly	Val	Pro	Val	Cys	His	Thr	Cys	Gly	Glu	His	Val
1					5					10				15	
Gly	Leu	Asn	Val	Asn	Gly	Glu	Pro	Phe	Val	Ala	Cys	His	Glu	Cys	Asn

[illegible]

	Met	Lys	Arg	Asp	Tyr	Glu	Glu	Tyr	Lys	Ile	Arg	Ile	Asn	Ala	Leu	Val	
				355					360					365			
5	Ala	Lys	Ala	Gln	Lys	Thr	Pro	Glu	Glu	Gly	Trp	Thr	Met	Gln	Asp	Gly	
			370				375					380					
	Thr	Pro	Trp	Pro	Gly	Asn	Asn	Pro	Arg	Asp	His	Pro	Gly	Met	Ile	Gln	
	385					390				395					400		
10	Val	Phe	Leu	Gly	Tyr	Ser	Gly	Ala	His	Asp	Ile	Glu	Gly	Asn	Glu	Leu	
				405					410					415			
	Pro	Arg	Leu	Val	Tyr	Val	Ser	Arg	Glu	Lys	Arg	Pro	Gly	Tyr	Gln	His	
				420					425					430			
15	His	Lys	Lys	Ala	Gly	Ala	Glu	Asn	Ala	Leu	Val	Arg	Val	Ser	Ala	Val	
			435					440					445				
	Leu	Thr	Asn	Ala	Pro	Phe	Ile	Leu	Asn	Leu	Asp	Cys	Asp	His	Tyr	Val	
		450					455				460						
20	Asn	Asn	Ser	Lys	Ala	Val	Arg	Glu	Ala	Met	Cys	Phe	Leu	Met	Asp	Pro	
	465					470				475					480		
	Gln	Val	Gly	Arg	Asp	Val	Cys	Tyr	Val	Gln	Phe	Pro	Gln	Arg	Phe	Asp	
				485					490					495			
25	Gly	Ile	Asp	Arg	Ser	Asp	Arg	Tyr	Ala	Asn	Arg	Asn	Thr	Val	Phe	Phe	
			500					505					510				
	Asp	Val	Asn	Met	Lys	Gly	Leu	Asp	Gly	Ile	Gln	Gly	Pro	Val	Tyr	Val	
		515					520				525						
30	Gly	Thr	Gly	Cys	Val	Phe	Asn	Arg	Gln	Ala	Leu	Tyr	Gly	Tyr	Gly	Pro	
		530				535				540							
	Pro	Ser	Met	Pro	Ser	Phe	Pro	Lys	Ser	Ser	Ser	Ser	Ser	Cys	Ser	Cys	
35	545					550				555				560			
	Cys	Cys	Pro	Gly	Lys	Lys	Glu	Pro	Lys	Asp	Pro	Ser	Glu	Leu	Tyr	Arg	
				565					570				575				
	Asp	Ala	Lys	Arg	Glu	Glu	Leu	Asp	Ala	Ala	Ile	Phe	Asn	Leu	Arg	Glu	
40				580					585				590				
	Ile	Asp	Asn	Tyr	Asp	Glu	Tyr	Glu	Arg	Ser	Met	Leu	Ile	Ser	Gln	Thr	
		595					600					605					
	Ser	Phe	Glu	Lys	Thr	Phe	Gly	Leu	Ser	Ser	Val	Phe	Ile	Glu	Ser	Thr	
45		610					615				620						
	Leu	Met	Glu	Asn	Gly	Gly	Val	Ala	Glu	Ser	Ala	Asn	Pro	Ser	Thr	Leu	
	625					630				635				640			
	Ile	Lys	Glu	Ala	Ile	His	Val	Ile	Gly	Cys	Gly	Tyr	Glu	Glu	Lys	Thr	
50				645					650				655				
	Ala	Trp	Gly	Lys	Glu	Ile	Gly	Trp	Ile	Tyr	Gly	Ser	Val	Thr	Glu	Asp	
			660					665				670					
55	Ile	Leu	Thr	Gly	Phe	Lys	Met	His	Cys	Arg	Gly	Trp	Arg	Ser	Ile	Tyr	

675 680 685
 Cys Met Pro Leu Arg Pro Ala Phe Lys Gly Ser Ala Pro Ile Asn Leu
 690 695 700
 5 Ser Asp Arg Leu His Gln Val Leu Arg Trp Ala Leu Gly Ser Val Glu
 705 710 715 720
 Ile Phe Leu Ser Arg His Cys Pro Leu Trp Tyr Gly Phe Gly Gly Gly
 725 730 735
 10 Arg Leu Lys Trp Leu Gln Arg Leu Ala Tyr Ile Asn Thr Ile Val Tyr
 740 745 750
 Pro Phe Thr Ser Leu Pro Leu Ile Ala Tyr Cys Ser Leu Pro Ala Ile
 755 760 765
 15 Cys Leu Leu Thr Gly Lys Phe Ile Ile Pro Thr Leu Ser Asn Leu Ala
 770 775 780
 Ser Val Leu Phe Leu Gly Leu Phe Leu Ser Ile Ile Val Thr Ala Val
 785 790 795 800
 Leu Glu Leu Arg Trp Ser Gly Val Ser Ile Glu Asp Leu Trp Arg Asn
 805 810 815
 25 Glu Gln Phe Trp Val Ile Gly Gly Val Ser Ala His Leu Phe Ala Val
 820 825 830
 Phe Gln Gly Phe Leu Lys Met Leu Ala Gly Ile Asp Thr Asn Phe Thr
 835 840 845
 30 Val Thr Ala Lys Ala Ala Asp Asp Ala Asp Phe Gly Glu Leu Tyr Ile
 850 855 860
 Val Lys Trp Thr Thr Leu Leu Ile Pro Pro Thr Thr Leu Leu Ile Val
 865 870 875 880
 35 Asn Met Val Gly Val Val Ala Gly Phe Ser Asp Ala Leu Asn Lys Gly
 885 890 895
 Tyr Glu Ala Trp Gly Pro Leu Phe Gly Lys Val Phe Phe Ser Phe Trp
 900 905 910
 40 Val Ile Leu His Leu Tyr Pro Phe Leu Lys Gly Leu Met Gly Arg Gln
 915 920 925
 Asn Arg Thr Pro Thr Ile Val Val Leu Trp Ser Val Leu Leu Ala Ser
 930 935 940
 45 Val Phe Ser Leu Val Trp Val Arg Ile Asn Pro Phe Val Ser Thr Ala
 945 950 955 960
 Asp Ser Thr Thr Val Ser Gln Ser Cys Ile Ser Ile Asp Cys
 965 970
 50

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3311 base pairs

(B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Gossypium hirsutum* L.

(C) INDIVIDUAL ISOLATE: Coker312

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 23..3142

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

15	CTTGGTCTCT TTTGGTTTTC CC ATG GCT TCA ACC ACC ATG GGC GCT GGC TTT	52
	Met Ala Ser Thr Thr Met Ala Ala Gly Phe	
	1 5 10	
20	GGT TCA CTT GCT GTT GAC GAG AAT OGG GGA TCA TCG ACA CAT CAA TCA	100
	Gly Ser Leu Ala Val Asp Glu Asn Arg Gly Ser Ser Thr His Gln Ser	
	15 20 25	
25	TCA ACG AAA ATA TGC AGG GTG TGT GGG GAT AAG ATC GGG CAA AAG GAA	148
	Ser Thr Lys Ile Cys Arg Val Cys Gly Asp Lys Ile Gly Gln Lys Glu	
	30 35 40	
30	AAC GGA CAA OCG TTC GTG GCT TGT CAT GTC TGT GCT TTC OCG GTT TGC	196
	Asn Gly Gln Pro Phe Val Ala Cys His Val Cys Ala Phe Pro Val Cys	
	45 50 55	
35	OGT OCT TGT TAT GAA TAT GAA AGG AGT GAA GGA AAC CAG TGC TGT OCT	244
	Arg Pro Cys Tyr Glu Tyr Glu Arg Ser Glu Gly Asn Gln Cys Cys Pro	
	60 65 70	
40	CAG TGC AAT ACT OGC TAT AAG OGT CAC AAA GGT AGT OCA AGA ATT TCA	292
	Gln Cys Asn Thr Arg Tyr Lys Arg His Lys Gly Ser Pro Arg Ile Ser	
	75 80 85 90	
45	GGG GAT GAA GAA GAT GAT TCA GAT CAA GAT GAT TTT GAT GAT GAA TTT	340
	Gly Asp Glu Glu Asp Asp Ser Asp Gln Asp Asp Phe Asp Asp Glu Phe	
	95 100 105	
50	CAG ATT AAG AAC OGC AAG GAT GAC TCC CAT OCA CAA CAT GAA AAT GAG	388
	Gln Ile Lys Asn Arg Lys Asp Asp Ser His Pro Gln His Glu Asn Glu	
	110 115 120	
55	GAA TAT AAT AAT AAT AAT CAT CAA TGG CAT CCC AAT GGT CAA GCT TTC	436
	Glu Tyr Asn Asn Asn Asn His Gln Trp His Pro Asn Gly Gln Ala Phe	
	125 130 135	
60	TCA GTT GGC GGA AGC ACG GCG GGG AAG GAT TTG GAA GGG GAT AAA GAG	484
	Ser Val Ala Gly Ser Thr Ala Gly Lys Asp Leu Glu Gly Asp Lys Glu	
	140 145 150	

	ATT	TAC	GGA	AGC	GAA	GAA	TGG	AAA	GAA	AGA	GTT	GAG	AAA	TGG	AAA	GTC	532
	Ile	Tyr	Gly	Ser	Glu	Glu	Trp	Lys	Glu	Arg	Val	Glu	Lys	Trp	Lys	Val	
5	155					160					165					170	
	AGG	CAA	GAA	AAA	AGA	GGT	TTG	GTA	AGC	AAC	GAT	AAT	GGC	GGA	AAT	GAT	580
	Arg	Gln	Glu	Lys	Arg	Gly	Leu	Val	Ser	Asn	Asp	Asn	Gly	Gly	Asn	Asp	
					175					180						185	
10	OCT	OCT	GAA	GAA	GAT	GAT	TAT	CTC	TTG	GCT	GAA	GCT	CGC	CAG	OCT	CTA	628
	Pro	Pro	Glu	Glu	Asp	Asp	Tyr	Leu	Leu	Ala	Glu	Ala	Arg	Gln	Pro	Leu	
				190					195					200			
	TGG	OGA	AAA	GTG	OCA	ATT	TOG	TCA	AGT	CTG	ATA	AGC	OCT	TAC	CGG	ATA	676
15	Trp	Arg	Lys	Val	Pro	Ile	Ser	Ser	Ser	Leu	Ile	Ser	Pro	Tyr	Arg	Ile	
			205					210					215				
	GTC	ATC	GTC	CTC	OGA	TTC	TTC	ATC	CTC	GCA	TTT	TTC	CTC	OGG	TTC	CGT	724
	Val	Ile	Val	Leu	Arg	Phe	Phe	Ile	Leu	Ala	Phe	Phe	Leu	Arg	Phe	Arg	
		220					225					230					
20	ATT	CTA	ACA	CCC	GCC	TAC	GAC	GCT	TAC	COG	TTA	TGG	CTA	ATC	TCT	GTC	772
	Ile	Leu	Thr	Pro	Ala	Tyr	Asp	Ala	Tyr	Pro	Leu	Trp	Leu	Ile	Ser	Val	
	235					240				245						250	
25	ATC	TGC	GAA	GTT	TGG	TTC	GCC	TTC	TOC	TGG	ATT	CTC	GAT	CAG	TTC	OCT	820
	Ile	Cys	Glu	Val	Trp	Phe	Ala	Phe	Ser	Trp	Ile	Leu	Asp	Gln	Phe	Pro	
				255					260					265			
	AAA	TGG	TTC	OCT	ATT	ACT	OGC	GAA	ACT	TAC	CTC	GAT	OGC	CTC	TOC	TTG	868
30	Lys	Trp	Phe	Pro	Ile	Thr	Arg	Glu	Thr	Tyr	Leu	Asp	Arg	Leu	Ser	Leu	
				270					275					280			
	AGG	TTC	GAA	OGT	GAA	GGA	GAG	CCC	AAT	CAA	CTT	GGC	CCC	GTC	GAC	GTC	916
	Arg	Phe	Glu	Arg	Glu	Gly	Glu	Pro	Asn	Gln	Leu	Gly	Pro	Val	Asp	Val	
35			285					290					295				
	TTC	GTC	AGT	AOC	GTT	GAC	CTT	CTC	AAG	GAA	CCC	CCC	ATC	ATA	ACC	GOC	964
	Phe	Val	Ser	Thr	Val	Asp	Leu	Leu	Lys	Glu	Pro	Pro	Ile	Ile	Thr	Ala	
		300					305					310					
40	AAC	GOG	GTT	CTA	TOG	ATC	TTG	GCC	GTC	GAT	TAC	COG	GTC	GAG	AAA	GTG	1012
	Asn	Ala	Val	Leu	Ser	Ile	Leu	Ala	Val	Asp	Tyr	Pro	Val	Glu	Lys	Val	
	315					320					325					330	
	TGT	TGT	TAT	GTG	TOG	GAC	GAT	GGT	GCT	TOC	ATG	CTT	CTT	TTC	GAT	TOG	1060
45	Cys	Cys	Tyr	Val	Ser	Asp	Asp	Gly	Ala	Ser	Met	Leu	Leu	Phe	Asp	Ser	
				335					340					345			
	TTG	TCT	GAA	ACG	GCT	GAG	TTC	GCG	AGG	AGA	TGG	GTT	COG	TTT	TGT	AAG	1108
	Leu	Ser	Glu	Thr	Ala												

	365	370	375	
	ATT GAT TAT TTG AAG GAC AAG GTC CAT OCT AGC TTT GTT AAA GAA OGG			1204
5	Ile Asp Tyr Leu Lys Asp Lys Val His Pro Ser Phe Val Lys Glu Arg			
	380	385	390	
	AGA GGC ATG AAA AGG GAA TAT GAA GAA TTT AAA GTA AGG ATC AAT GCA			1252
	Arg Ala Met Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg Ile Asn Ala			
10	395	400	405	410
	TTA GTA GCA AAA GCT CAG AAG AAA OCA GAA GAA GGA TGG GTG ATG CAA			1300
	Leu Val Ala Lys Ala Gln Lys Lys Pro Glu Glu Gly Trp Val Met Gln			
	415	420	425	
15	GAT GGC ACC OCA TGG CCC GGA AAT AAC ACT CGT GAT CAT OCT GGA ATG			1348
	Asp Gly Thr Pro Trp Pro Gly Asn Asn Thr Arg Asp His Pro Gly Met			
	430	435	440	
	ATT CAG GTC TAT CTA GGA AGT GGC GGT GCA CTC GAT GTG GAT GGC AAA			1396
20	Ile Gln Val Tyr Leu Gly Ser Ala Gly Ala Leu Asp Val Asp Gly Lys			
	445	450	455	
	GAG CTG OCT OGA CTT GTC TAT GTT TCT CGT GAG AAA OGA OCT GGT TAT			1444
	Glu Leu Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg Pro Gly Tyr			
	460	465	470	
25	CAG CAC CAT AAG AAA GGC GGT GCT GAG AAT GCT CTG GTT OGA GTT TCT			1492
	Gln His His Lys Lys Ala Gly Ala Glu Asn Ala Leu Val Arg Val Ser			
	475	480	485	490
	GCA GTG CTT ACT AAT GCA CCC TTC ATA TTG AAT CTG GAT TGT GAT CAT			1540
30	Ala Val Leu Thr Asn Ala Pro Phe Ile Leu Asn Leu Asp Cys Asp His			
	495	500	505	
	TAC ATC AAC AAT AGC AAG GGC ATG AGG GAA GCG ATG TGC TTT TTA ATG			1588
	Tyr Ile Asn Asn Ser Lys Ala Met Arg Glu Ala Met Cys Phe Leu Met			
35	510	515	520	
	GAT OCT CAG TTT GGA AAG AAG CTT TGT TAT GTT CAA TTT OCA CAG AGA			1636
	Asp Pro Gln Phe Gly Lys Lys Leu Cys Tyr Val Gln Phe Pro Gln Arg			
	525	530	535	
40	TTT GAT GGT ATT GAT CGT CAT GAT OGA TAT GCT AAT OGA AAT GTT GTC			1684
	Phe Asp Gly Ile Asp Arg His Asp Arg Tyr Ala Asn Arg Asn Val Val			
	540	545	550	
	TTC TTT GAT ATC AAC ATG TTG GGA TTA GAT GGA CTT CAA GGC OCT GTA			1732
45	Phe Phe Asp Ile Asn Met Leu Gly Leu Asp Gly Leu Gln Gly Pro Val			
	555	560	565	570
	TAT GTA GGC ACA GGG TGT GTT TTC AAC AGG CAG GCA TTG TAT GGC TAC			1780
	Tyr Val Gly Thr Gly Cys Val Phe Asn Arg Gln Ala Leu Tyr Gly Tyr			
50	575	580	585	
	GAT CCA CCA GTC TCT GAG AAA OGA CCA AAG ATG ACA TGT GAT TGC TGG			1828

	Asp	Pro	Pro	Val	Ser	Glu	Lys	Arg	Pro	Lys	Met	Thr	Cys	Asp	Cys	Trp	
				590					595					600			
5	OCT	TCT	TGG	TGT	TGC	TGT	TGT	TGC	GGA	GGT	TCT	AGG	AAG	AAA	TCA	AAG	1876
	Pro	Ser	Trp	Cys	Cys	Cys	Cys	Cys	Gly	Gly	Ser	Arg	Lys	Lys	Ser	Lys	
			605					610					615				
	AAG	AAA	GGT	GAA	AAG	AAG	GGC	TTA	CTC	GGA	GGT	CTT	TTA	TAC	GGA	AAA	1924
10	Lys	Lys	Gly	Glu	Lys	Lys	Gly	Leu	Leu	Gly	Gly	Leu	Leu	Tyr	Gly	Lys	
			620				625					630					
	AAG	AAG	AAG	ATG	ATG	GGC	AAA	AAC	TAT	GTG	AAA	AAA	GGG	TCT	GCA	OCA	1972
	Lys	Lys	Lys	Met	Met	Gly	Lys	Asn	Tyr	Val	Lys	Lys	Gly	Ser	Ala	Pro	
15	635					640				645					650		
	GTC	TTT	GAT	CTC	GAA	GAA	ATC	GAA	GAA	GGG	CTT	GAA	GGA	TAC	GAA	GAA	2020
	Val	Phe	Asp	Leu	Glu	Glu	Ile	Glu	Glu	Gly	Leu	Glu	Gly	Tyr	Glu	Glu	
				655				660					665				
20	TTG	GAG	AAA	TOG	ACA	TTA	ATG	TOG	CAG	AAG	AAT	TTC	GAG	AAA	OGA	TTC	2068
	Leu	Glu	Lys	Ser	Thr	Leu	Met	Ser	Gln	Lys	Asn	Phe	Glu	Lys	Arg	Phe	
				670				675				680					
	GGA	CAA	TCA	COG	GTT	TTC	ATT	GOC	TCA	ACT	TTG	ATG	GAA	AAT	GGT	GGC	2116
25	Gly	Gln	Ser	Pro	Val	Phe	Ile	Ala	Ser	Thr	Leu	Met	Glu	Asn	Gly	Gly	
			685				690				695						
	CTT	OCT	GAA	GGA	ACT	AAT	TOC	ACA	TCA	CTG	ATT	AAA	GAG	GCC	ATT	CAC	2164
	Leu	Pro	Glu	Gly	Thr	Asn	Ser	Thr	Ser	Leu	Ile	Lys	Glu	Ala	Ile	His	
30			700				705				710						
	GTA	ATT	AGC	TGT	GGT	TAT	GAA	GAA	AAA	ACT	GAG	TGG	GGC	AAA	GAG	ATC	2212
	Val	Ile	Ser	Cys	Gly	Tyr	Glu	Glu	Lys	Thr	Glu	Trp	Gly	Lys	Glu	Ile	
			715			720				725			730				
35	GGA	TGG	ATT	TAT	GGG	TOG	GTG	ACG	GAA	GAT	ATA	TTA	ACA	GGT	TTC	AAG	2260
	Gly	Trp	Ile	Tyr	Gly	Ser	Val	Thr	Glu	Asp	Ile	Leu	Thr	Gly	Phe	Lys	
			735				740			745							
	ATG	CAT	TGT	AGA	GGG	TGG	AAA	TOG	GTT	TAT	TGT	GTA	COG	AAA	AGA	COG	2308
40	Met	His	Cys	Arg	Gly	Trp	Lys	Ser	Val	Tyr	Cys	Val	Pro	Lys	Arg	Pro	
			750				755				760						
	GCA	TTC	AAA	GGG	TOC	GCT	CCA	ATC	AAT	CTC	TOG	GAT	OGG	TTG	CAC	CAA	2356
	Ala	Phe	Lys	Gly	Ser	Ala	Pro	Ile	Asn	Leu	Ser	Asp	Arg	Leu	His	Gln	
45			765				770				775						
	GTT	TTG	AGA	TGG	GCA	CTT	GGT	TCT	GTA	GAA	ATT	TTC	CTT	AGT	CGT	CAC	2404
	Val	Leu	Arg	Trp	Ala	Leu	Gly	Ser	Val	Glu	Ile	Phe	Leu	Ser	Arg	His	
			780			785				790							
50	TGT	CCA	CTT	TGG	TAT	GGT	TAT	GGT	GGA	AAA	CTG	AAA	TGG	CTC	GAG	AGG	2452
	Cys	Pro	Leu	Trp	Tyr	Gly	Tyr	Gly	Gly	Lys	Leu	Lys	Trp	Leu	Glu	Arg	
			795			800				805					810		

55

	CTT	GCT	TAT	ATC	AAC	ACC	ATT	GTT	TAC	OCT	TTC	AOC	TOG	ATC	OCT	TTA	2500
	Leu	Ala	Tyr	Ile	Asn	Thr	Ile	Val	Tyr	Pro	Phe	Thr	Ser	Ile	Pro	Leu	
5				815					820						825		
	CTC	GOC	TAT	TGT	ACT	ATT	OCA	GCT	GTT	TGT	CTT	CTC	ACC	GGC	AAA	TTC	2548
	Leu	Ala	Tyr	Cys	Thr	Ile	Pro	Ala	Val	Cys	Leu	Leu	Thr	Gly	Lys	Phe	
				830					835						840		
10	ATC	ATT	OCA	ACT	CTA	AGC	AAC	CTT	ACA	AGT	GTG	TGG	TTC	TTG	GCA	CTT	2596
	Ile	Ile	Pro	Thr	Leu	Ser	Asn	Leu	Thr	Ser	Val	Trp	Phe	Leu	Ala	Leu	
				845					850						855		
	TTC	CTC	TOC	ATC	ATT	GCA	ACT	GGA	GTG	CTT	GAA	CTT	OGA	TGG	AGC	GGG	2644
15	Phe	Leu	Ser	Ile	Ile	Ala	Thr	Gly	Val	Leu	Glu	Leu	Arg	Trp	Ser	Gly	
				860					865					870			
	GTT	AGC	ATC	CAA	GAC	TGG	TGG	OGC	AAT	GAA	CAA	TTC	TGG	GTG	ATC	GGA	2692
	Val	Ser	Ile	Gln	Asp	Trp	Trp	Arg	Asn	Glu	Gln	Phe	Trp	Val	Ile	Gly	
20				875			880				885					890	
	GGT	GTC	TOC	GOC	CAT	CTT	TTT	GCT	GTC	TTC	CAG	GGC	CTC	CTC	AAA	GTC	2740
	Gly	Val	Ser	Ala	His	Leu	Phe	Ala	Val	Phe	Gln	Gly	Leu	Leu	Lys	Val	
				895					900						905		
25	CTA	GCT	GGA	GTA	GAC	ACC	AAC	TTC	ACC	GTA	ACA	GCA	AAA	GCA	GCA	GAC	2788
	Leu	Ala	Gly	Val	Asp	Thr	Asn	Phe	Thr	Val	Thr	Ala	Lys	Ala	Ala	Asp	
				910					915					920			
	GAT	ACA	GAA	TTC	GGT	GAA	CTT	TAT	CTC	TTC	AAA	TGG	ACA	ACT	CTC	TTA	2836
30	Asp	Thr	Glu	Phe	Gly	Glu	Leu	Tyr	Leu	Phe	Lys	Trp	Thr	Thr	Leu	Leu	
				925					930					935			
	ATC	OCT	OOC	ACA	ACT	CTG	ATA	ATA	CTG	AAC	ATG	GTC	GGA	GTC	GTG	GOC	2884
	Ile	Pro	Pro	Thr	Thr	Leu	Ile	Ile	Leu	Asn	Met	Val	Gly	Val	Val	Ala	
35				940				945					950				
	GGA	GTT	TCA	GAC	GCA	ATC	AAC	AAC	GGC	TAT	GGT	TCA	TGG	GGT	OCA	TTG	2932
	Gly	Val	Ser	Asp	Ala	Ile	Asn	Asn	Gly	Tyr	Gly	Ser	Trp	Gly	Pro	Leu	
				955			960				965				970		
40	TTC	GGC	AAA	CTG	TTC	TTC	GCA	TTC	TGG	GTC	ATT	CTT	CAT	CTT	TAC	OCA	2980
	Phe	Gly	Lys	Leu	Phe	Phe	Ala	Phe	Trp	Val	Ile	Leu	His	Leu	Tyr	Pro	
				975					980						985		
	TTC	CTC	AAA	GGT	TTG	ATG	GGG	AGA	CAA	AAC	AGG	ACG	OCC	AOC	ATT	GTT	3028
45	Phe	Leu	Lys	Gly	Leu	Met	Gly	Arg	Gln	Asn	Arg	Thr	Pro	Thr	Ile	Val	
				990					995						1000		
	GTG	CTT	TGG	TOC	ATA	CTT	TTG	GCA	TOG	ATT	TTC	TCA	CTG	GTT	TGG	GTA	3076
	Val	Leu	Trp	Ser	Ile	Leu	Leu	Ala	Ser	Ile	Phe	Ser	Leu	Val	Trp	Val	
				1005					1010					1015			
50	OGG	ATC	GAT	OCC	TTC	TTG	OCC	AAA	CAA	ACA	GGT	OCA	GTT	CTT	AAA	CAA	3124
	Arg	Ile	Asp	Pro	Phe	Leu	Pro	Lys	Gln	Thr	Gly	Pro	Val	Leu	Lys	Gln	

55

1020 1025 1030
 TGT GGC GTG GAG TGC TAAATGGTGT TTTACAAACC TTCTTATTA TTTTATTTTC 3179
 Cys Gly Val Glu Cys
 1035
 CCTTTTGGC ACTACTGTG ATTTGCTGTG ATTCTAAAAG GGATTATCT TGTGTGTAAG 3239
 AAGTCTCTTA TGATTTTGTG GGTTCATTT AATTTCTATA TGGTAAAAAA ATATTTCTTT 3299
 AAATTAAC TA 3311

(2) INFORMATION FOR SEQ ID NO: 4:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1039 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(11) MOLECULE TYPE: protein

(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Met Ala Ser Thr Thr Met Ala Ala Gly Phe Gly Ser Leu Ala Val Asp
 1 5 10 15
 Glu Asn Arg Gly Ser Ser Thr His Gln Ser Ser Thr Lys Ile Cys Arg
 20 25 30
 Val Cys Gly Asp Lys Ile Gly Gln Lys Glu Asn Gly Gln Pro Phe Val
 35 40 45
 Ala Cys His Val Cys Ala Phe Pro Val Cys Arg Pro Cys Tyr Glu Tyr
 50 55 60
 Glu Arg Ser Glu Gly Asn Gln Cys Cys Pro Gln Cys Asn Thr Arg Tyr
 65 70 75 80
 Lys Arg His Lys Gly Ser Pro Arg Ile Ser Gly Asp Glu Glu Asp Asp
 85 90 95
 Ser Asp Gln Asp Asp Phe Asp Asp Glu Phe Gln Ile Lys Asn Arg Lys
 100 105 110
 Asp Asp Ser His Pro Gln His Glu Asn Glu Glu Tyr Asn Asn Asn Asn
 115 120 125
 His Gln Trp His Pro Asn Gly Gln Ala Phe Ser Val Ala Gly Ser Thr
 130 135 140
 Ala Gly Lys Asp Leu Glu Gly Asp Lys Glu Ile Tyr Gly Ser Glu Glu
 145 150 155 160
 Trp Lys Glu Arg Val Glu Lys Trp Lys Val Arg Gln Glu Lys Arg Gly
 165 170 175
 Leu Val Ser Asn Asp Asn Gly Gly Asn Asp Pro Pro Glu Glu Asp Asp
 180 185 190
 Tyr Leu Leu Ala Glu Ala Arg Gln Pro Leu Trp Arg Lys Val Pro Ile
 195 200 205

	Ser	Ser	Ser	Leu	Ile	Ser	Pro	Tyr	Arg	Ile	Val	Ile	Val	Leu	Arg	Phe	
	210						215					220					
5	Phe	Ile	Leu	Ala	Phe	Phe	Leu	Arg	Phe	Arg	Ile	Leu	Thr	Pro	Ala	Tyr	
	225					230					235					240	
	Asp	Ala	Tyr	Pro	Leu	Trp	Leu	Ile	Ser	Val	Ile	Cys	Glu	Val	Trp	Phe	
					245					250					255		
10	Ala	Phe	Ser	Trp	Ile	Leu	Asp	Gln	Phe	Pro	Lys	Trp	Phe	Pro	Ile	Thr	
				260					265					270			
	Arg	Glu	Thr	Tyr	Leu	Asp	Arg	Leu	Ser	Leu	Arg	Phe	Glu	Arg	Glu	Gly	
			275				280						285				
15	Glu	Pro	Asn	Gln	Leu	Gly	Pro	Val	Asp	Val	Phe	Val	Ser	Thr	Val	Asp	
	290					295					300						
	Leu	Leu	Lys	Glu	Pro	Pro	Ile	Ile	Thr	Ala	Asn	Ala	Val	Leu	Ser	Ile	
	305					310					315					320	
20	Leu	Ala	Val	Asp	Tyr	Pro	Val	Glu	Lys	Val	Cys	Cys	Tyr	Val	Ser	Asp	
				325						330					335		
	Asp	Gly	Ala	Ser	Met	Leu	Leu	Phe	Asp	Ser	Leu	Ser	Glu	Thr	Ala	Glu	
				340					345					350			
25	Phe	Ala	Arg	Arg	Trp	Val	Pro	Phe	Cys	Lys	Lys	His	Asn	Val	Glu	Pro	
			355				360						365				
	Arg	Ala	Pro	Glu	Phe	Tyr	Phe	Asn	Glu	Lys	Ile	Asp	Tyr	Leu	Lys	Asp	
	370					375					380						
30	Lys	Val	His	Pro	Ser	Phe	Val	Lys	Glu	Arg	Arg	Ala	Met	Lys	Arg	Glu	
	385					390					395					400	
	Tyr	Glu	Glu	Phe	Lys	Val	Arg	Ile	Asn	Ala	Leu	Val	Ala	Lys	Ala	Gln	
				405						410				415			
35	Lys	Lys	Pro	Glu	Glu	Gly	Trp	Val	Met	Gln	Asp	Gly	Thr	Pro	Trp	Pro	
			420						425				430				
	Gly	Asn	Asn	Thr	Arg	Asp	His	Pro	Gly	Met	Ile	Gln	Val	Tyr	Leu	Gly	
		435				440					445						
40	Ser	Ala	Gly	Ala	Leu	Asp	Val	Asp	Gly	Lys	Glu	Leu	Pro	Arg	Leu	Val	
	450					455					460						
	Tyr	Val	Ser	Arg	Glu	Lys	Arg	Pro	Gly	Tyr	Gln	His	His	Lys	Lys	Ala	
45	465					470					475					480	
	Gly	Ala	Glu	Asn	Ala	Leu	Val	Arg	Val	Ser	Ala	Val	Leu	Thr	Asn	Ala	
				485						490				495			
	Pro	Phe	Ile	Leu	Asn	Leu	Asp	Cys	Asp	His	Tyr	Ile	Asn	Asn	Ser	Lys	
50				500					505				510				
	Ala	Met	Arg	Glu	Ala	Met	Cys	Phe	Leu	Met	Asp	Pro	Gln	Phe	Gly	Lys	
		515						520					525				
55	Lys	Leu	Cys	Tyr	Val	Gln	Phe	Pro	Gln	Arg	Phe	Asp	Gly	Ile	Asp	Arg	

	530	535	540
	His Asp Arg Tyr Ala Asn Arg Asn Val Val Phe Phe Asp Ile Asn Met		
5	545	550	555 560
	Leu Gly Leu Asp Gly Leu Gln Gly Pro Val Tyr Val Gly Thr Gly Cys		
	565	570	575
	Val Phe Asn Arg Gln Ala Leu Tyr Gly Tyr Asp Pro Pro Val Ser Glu		
10	580	585	590
	Lys Arg Pro Lys Met Thr Cys Asp Cys Trp Pro Ser Trp Cys Cys Cys		
	595	600	605
	Cys Cys Gly Gly Ser Arg Lys Lys Ser Lys Lys Lys Gly Glu Lys Lys		
15	610	615	620
	Gly Leu Leu Gly Gly Leu Leu Tyr Gly Lys Lys Lys Lys Met Met Gly		
	625	630	635 640
	Lys Asn Tyr Val Lys Lys Gly Ser Ala Pro Val Phe Asp Leu Glu Glu		
20	645	650	655
	Ile Glu Glu Gly Leu Glu Gly Tyr Glu Glu Leu Glu Lys Ser Thr Leu		
	660	665	670
	Met Ser Gln Lys Asn Phe Glu Lys Arg Phe Gly Gln Ser Pro Val Phe		
25	675	680	685
	Ile Ala Ser Thr Leu Met Glu Asn Gly Gly Leu Pro Glu Gly Thr Asn		
	690	695	700
	Ser Thr Ser Leu Ile Lys Glu Ala Ile His Val Ile Ser Cys Gly Tyr		
30	705	710	715 720
	Glu Glu Lys Thr Glu Trp Gly Lys Glu Ile Gly Trp Ile Tyr Gly Ser		
	725	730	735
	Val Thr Glu Asp Ile Leu Thr Gly Phe Lys Met His Cys Arg Gly Trp		
35	740	745	750
	Lys Ser Val Tyr Cys Val Pro Lys Arg Pro Ala Phe Lys Gly Ser Ala		
	755	760	765
	Pro Ile Asn Leu Ser Asp Arg Leu His Gln Val Leu Arg Trp Ala Leu		
40	770	775	780
	Gly Ser Val Glu Ile Phe Leu Ser Arg His Cys Pro Leu Trp Tyr Gly		
	785	790	795 800
	Tyr Gly Gly Lys Leu Lys Trp Leu Glu Arg Leu Ala Tyr Ile Asn Thr		
45	805	810	815
	Ile Val Tyr Pro Phe Thr Ser Ile Pro Leu Leu Ala Tyr Cys Thr Ile		
	820	825	830
	Pro Ala Val Cys Leu Leu Thr Gly Lys Phe Ile Ile Pro Thr Leu Ser		
50	835	840	845
	Asn Leu Thr Ser Val Trp Phe Leu Ala Leu Phe Leu Ser Ile Ile Ala		
	850	855	860
55			

Thr Gly Val Leu Glu Leu Arg Trp Ser Gly Val Ser Ile Gln Asp Trp
 865 870 875 880
 Trp Arg Asn Glu Gln Phe Trp Val Ile Gly Gly Val Ser Ala His Leu
 885 890 895
 Phe Ala Val Phe Gln Gly Leu Leu Lys Val Leu Ala Gly Val Asp Thr
 900 905 910
 Asn Phe Thr Val Thr Ala Lys Ala Ala Asp Asp Thr Glu Phe Gly Glu
 915 920 925
 Leu Tyr Leu Phe Lys Trp Thr Thr Leu Leu Ile Pro Pro Thr Thr Leu
 930 935 940
 Ile Ile Leu Asn Met Val Gly Val Val Ala Gly Val Ser Asp Ala Ile
 945 950 955 960
 Asn Asn Gly Tyr Gly Ser Trp Gly Pro Leu Phe Gly Lys Leu Phe Phe
 965 970 975
 Ala Phe Trp Val Ile Leu His Leu Tyr Pro Phe Leu Lys Gly Leu Met
 980 985 990
 Gly Arg Gln Asn Arg Thr Pro Thr Ile Val Val Leu Trp Ser Ile Leu
 995 1000 1005
 Leu Ala Ser Ile Phe Ser Leu Val Trp Val Arg Ile Asp Pro Phe Leu
 1010 1015 1020
 Pro Lys Gln Thr Gly Pro Val Leu Lys Gln Cys Gly Val Glu Cys
 1025 1030 1035

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2033 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Gossypium hirsutum* L.
- (C) INDIVIDUAL ISOLATE: Coker312

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1857

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

CCG ACA TTC GTG AAG GAG CGT OGA GCT ATG AAG AGA GAA TAT GAA GAA 48
 Pro Thr Phe Val Lys Glu Arg Arg Ala Met Lys Arg Glu Tyr Glu Glu
 1 5 10 15
 TTC AAG GTT AGG ATA AAT GCA CTT GTA GCC AAA GCC CAA AAG GTT CCT 96

	Phe Lys Val Arg Ile Asn Ala Leu Val Ala Lys Ala Gln Lys Val Pro	
	20 25 30	
5	CCA GAA GGG TGG ATC ATG CAA GAT GGG ACA OCA TGG OCA GGA AAC AAT	144
	Pro Glu Gly Trp Ile Met Gln Asp Gly Thr Pro Trp Pro Gly Asn Asn	
	35 40 45	
	ACT AAA GAT CAC OCT GGT ATG ATT CAA GTA TTT CTC GGT CAA AGT GGA	192
10	Thr Lys Asp His Pro Gly Met Ile Gln Val Phe Leu Gly Gln Ser Gly	
	50 55 60	
	GGC CAT GAT ACC GAA GGA AAT GAG CTT OCT CGT CTC GTC TAT GTA TCT	240
	Gly His Asp Thr Glu Gly Asn Glu Leu Pro Arg Leu Val Tyr Val Ser	
15	65 70 75 80	
	CGA GAG AAA AGG OCT GGT TTC TTG CAT CAC AAG AAA GCT GGT GGC ATG	288
	Arg Glu Lys Arg Pro Gly Phe Leu His His Lys Lys Ala Gly Ala Met	
	85 90 95	
20	AAC GGC CTT GTT CCG GTC TCG GGG GTG CTC ACA AAT GCT OCT TTT ATG	336
	Asn Ala Leu Val Arg Val Ser Gly Val Leu Thr Asn Ala Pro Phe Met	
	100 105 110	
	TTG AAC TTG GAT TGT GAC CAT TAT TTA AAT AAC AGC AAG GCT GTA AGA	384
25	Leu Asn Leu Asp Cys Asp His Tyr Leu Asn Asn Ser Lys Ala Val Arg	
	115 120 125	
	GAG GCT ATG TGT TTC TTG ATG GAC OCT CAA ATT GGA AGA AAG GTT TGC	432
	Glu Ala Met Cys Phe Leu Met Asp Pro Gln Ile Gly Arg Lys Val Cys	
30	130 135 140	
	TAT GTC CAA TTC OCT CAA CGT TTC GAT GGT ATT GAT AGA CAT GAT OGA	480
	Tyr Val Gln Phe Pro Gln Arg Phe Asp Gly Ile Asp Arg His Asp Arg	
	145 150 155 160	
35	TAT GGC AAT CCG AAC ACA GTT TTC TTT GAT ATT AAC ATG AAA GGT CTA	528
	Tyr Ala Asn Arg Asn Thr Val Phe Phe Asp Ile Asn Met Lys Gly Leu	
	165 170 175	
	GAT GGT ATA CAA GGC OCT GTA TAT GTC GGC ACG GGG TGT GTT TTC AGA	576
40	Asp Gly Ile Gln Gly Pro Val Tyr Val Gly Thr Gly Cys Val Phe Arg	
	180 185 190	
	AGG CAA GCT CTT TAT GGT TAT GAA OCT OCA AAG GGA OCT AAG GGC CCG	624
	Arg Gln Ala Leu Tyr Gly Tyr Glu Pro Pro Lys Gly Pro Lys Arg Pro	
45	195 200 205	
	AAA ATG GTA ACC TGT GGT TGC TGC OCT TGT TTT GGA GGC GGC AGA AAG	672
	Lys Met Val Thr Cys Gly Cys Cys Pro Cys Phe Gly Arg Arg Arg Lys	
	210 215 220	
50	GAC AAA AAG CAC TCT AAG GAT GGT GGA AAT GCA AAT GGT CTA AGC CTA	720
	Asp Lys Lys His Ser Lys Asp Gly Gly Asn Ala Asn Gly Leu Ser Leu	
	225 230 235 240	

55

	GAA GCA GCC AAA GAT GAC AAG GAG TTA TTG ATG TOC CAC ATG AAC TTT	768
	Glu Ala Ala Lys Asp Asp Lys Glu Leu Leu Met Ser His Met Asn Phe	
5	245 250 255	
	GAA AAG AAA TTT GGA CAA TCA GGC ATT TTT GTA ACT TCA ACA CTG ATG	816
	Glu Lys Lys Phe Gly Gln Ser Ala Ile Phe Val Thr Ser Thr Leu Met	
	260 265 270	
10	GAA CAA GGT GGT GTC OCT OCT TCT TCA AGC CCC GCA GCT TTG CTC AAA	864
	Glu Gln Gly Gly Val Pro Pro Ser Ser Ser Pro Ala Ala Leu Leu Lys	
	275 280 285	
	GAA GGC ATT CAT GTA ATT AGT TGT GGT TAT GAA GAC AAA ACA GAA TGG	912
15	Glu Ala Ile His Val Ile Ser Cys Gly Tyr Glu Asp Lys Thr Glu Trp	
	290 295 300	
	GGA AGC GAG CTT GGC TGG ATT TAC GGC TGG ATT ACA GAA GAT ATC TTA	960
	Gly Ser Glu Leu Gly Trp Ile Tyr Gly Ser Ile Thr Glu Asp Ile Leu	
	305 310 315 320	
20	ACA GGA TTC AAG ATG CAT TGC CGT GGA TGG AGA TCA ATA TAC TGC ATG	1008
	Thr Gly Phe Lys Met His Cys Arg Gly Trp Arg Ser Ile Tyr Cys Met	
	325 330 335	
	OCA AAG TTG OCT GCA TTC AAG GGT TCA GCT CCC ATC AAT CTA TCG GAT	1056
25	Pro Lys Leu Pro Ala Phe Lys Gly Ser Ala Pro Ile Asn Leu Ser Asp	
	340 345 350	
	CGT CTA AAC CAA GTC CTT CGA TGG GCA CTC GGT TCT GTT GAA ATT TTC	1104
	Arg Leu Asn Gln Val Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Phe	
30	355 360 365	
	TTT AGT CAT CAT TGC OCA GCA TGG TAT GGT TTC AAG GGA GGA AAG CTA	1152
	Phe Ser His His Cys Pro Ala Trp Tyr Gly Phe Lys Gly Gly Lys Leu	
	370 375 380	
35	AAA TGG CTT GAA CGA TTC GCA TAT GTC AAC ACA ACC ATC TAC CCC TTC	1200
	Lys Trp Leu Glu Arg Phe Ala Tyr Val Asn Thr Thr Ile Tyr Pro Phe	
	385 390 395 400	
	ACA TCT TTA OCA CTT CTC GGC TAT TGT ACC CTA CCG GCA ATC TGT TTA	1248
40	Thr Ser Leu Pro Leu Leu Ala Tyr Cys Thr Leu Pro Ala Ile Cys Leu	
	405 410 415	
	CTT ACC GAT AAA TTT ATC ATG OCA CCG ATA AGC ACC TTT GCA AGT CTA	1296
	Leu Thr Asp Lys Phe Ile Met Pro Pro Ile Ser Thr Phe Ala Ser Leu	
	420 425 430	
45	TTC TTC ATT GGC TTG TTT CTT TCA ATC TTT GCA ACT GGT ATT CTC GAG	1344
	Phe Phe Ile Ala Leu Phe Leu Ser Ile Phe Ala Thr Gly Ile Leu Glu	
	435 440 445	
50	CTA AGG TGG AGT GGA GTA AGC ATT GAA GAA TGG TGG AGG AAT GAG CAA	1392
	Leu Arg Trp Ser Gly Val Ser Ile Glu Glu Trp Trp Arg Asn Glu Gln	

	450	455	460	
	TTT TGG GTC ATC GGT GGC ATT TCG GCA CAT TTG TTC GCT GTT ATC CAA			1440
5	Phe Trp Val Ile Gly Gly Ile Ser Ala His Leu Phe Ala Val Ile Gln			
	465	470	475	480
	GGC TTG TTG AAA GTT CTA GCT GGT ATT GAC ACT AAT TTC ACT GTC ACA			1488
	Gly Leu Leu Lys Val Leu Ala Gly Ile Asp Thr Asn Phe Thr Val Thr			
		485	490	495
10	TOC AAG GCA ACT GAT GAC GAG GAG TTC GGG GAA TTG TAT ACT TTC AAA			1536
	Ser Lys Ala Thr Asp Asp Glu Glu Phe Gly Glu Leu Tyr Thr Phe Lys			
		500	505	510
15	TGG ACA ACC CTT CTA ATT OCT OCT ACT ACC GTC TTA ATC ATC AAT TTA			1584
	Trp Thr Thr Leu Leu Ile Pro Pro Thr Thr Val Leu Ile Ile Asn Leu			
		515	520	525
	GTC GGT GTC GTT GCA GGC ATC TCG GAT GCC ATA AAC AAT GGA TAC CAA			1632
20	Val Gly Val Val Ala Gly Ile Ser Asp Ala Ile Asn Asn Gly Tyr Gln			
		530	535	540
	TCA TGG GGA OCT CTT TTT GGG AAG CTC TTC TTC TCT TTC TGG GTG ATT			1680
	Ser Trp Gly Pro Leu Phe Gly Lys Leu Phe Phe Ser Phe Trp Val Ile			
		545	550	555
25	GTC CAT CTC TAT CCA TTC CTC AAA GGT TTA ATG GGG AGA CAA AAC OGG			1728
	Val His Leu Tyr Pro Phe Leu Lys Gly Leu Met Gly Arg Gln Asn Arg			
		565	570	575
30	ACA CCA ACC ATT GTT GTT ATA TGG TCA GTG CTA TTG GCT TCA ATC TTC			1776
	Thr Pro Thr Ile Val Val Ile Trp Ser Val Leu Leu Ala Ser Ile Phe			
		580	585	590
	TOC TTG CTT TGG GTC CGA ATT GAT CCA TTT GTG ATG AAA ACC AAA GGA			1824
	Ser Leu Leu Trp Val Arg Ile Asp Pro Phe Val Met Lys Thr Lys Gly			
35		595	600	605
	CCA GAC ACT ACA ATG TGT GGC ATT AAC TGT TGAAAAAAA TCATCTTGOG			1874
	Pro Asp Thr Thr Met Cys Gly Ile Asn Cys			
		610	615	
40	TGGTTCCTTT AGATTATGGT ATGTGATGTA TGAACAAACA AGAATGGAGA TGCACAAGAC			1934
	AGAATAAAAT TAGAGTGAAA GTTTGTGTGTA GTTATATATT CATTCTACCA ACTATAAGTT			1994
	TTGTCATTCA ATTGAAAATA GCTCAACTTT GTGATCAAA			2033

(2) INFORMATION FOR SEQ ID NO: 6:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 618 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(11) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: C-terminal fragment

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

5	Pro	Thr	Phe	Val	Lys	Glu	Arg	Arg	Ala	Met	Lys	Arg	Glu	Tyr	Glu	Glu	1	5	10	15
	Phe	Lys	Val	Arg	Ile	Asn	Ala	Leu	Val	Ala	Lys	Ala	Gln	Lys	Val	Pro	20	25	30	
10	Pro	Glu	Gly	Trp	Ile	Met	Gln	Asp	Gly	Thr	Pro	Trp	Pro	Gly	Asn	Asn	35	40	45	
	Thr	Lys	Asp	His	Pro	Gly	Met	Ile	Gln	Val	Phe	Leu	Gly	Gln	Ser	Gly	50	55	60	
15	Gly	His	Asp	Thr	Glu	Gly	Asn	Glu	Leu	Pro	Arg	Leu	Val	Tyr	Val	Ser	65	70	75	80
	Arg	Glu	Lys	Arg	Pro	Gly	Phe	Leu	His	His	Lys	Lys	Ala	Gly	Ala	Met	85	90	95	
20	Asn	Ala	Leu	Val	Arg	Val	Ser	Gly	Val	Leu	Thr	Asn	Ala	Pro	Phe	Met	100	105	110	
	Leu	Asn	Leu	Asp	Cys	Asp	His	Tyr	Leu	Asn	Asn	Ser	Lys	Ala	Val	Arg	115	120	125	
25	Glu	Ala	Met	Cys	Phe	Leu	Met	Asp	Pro	Gln	Ile	Gly	Arg	Lys	Val	Cys	130	135	140	
	Tyr	Val	Gln	Phe	Pro	Gln	Arg	Phe	Asp	Gly	Ile	Asp	Arg	His	Asp	Arg	145	150	155	160
30	Tyr	Ala	Asn	Arg	Asn	Thr	Val	Phe	Phe	Asp	Ile	Asn	Met	Lys	Gly	Leu	165	170	175	
	Asp	Gly	Ile	Gln	Gly	Pro	Val	Tyr	Val	Gly	Thr	Gly	Cys	Val	Phe	Arg	180	185	190	
35	Arg	Gln	Ala	Leu	Tyr	Gly	Tyr	Glu	Pro	Pro	Lys	Gly	Pro	Lys	Arg	Pro	195	200	205	
	Lys	Met	Val	Thr	Cys	Gly	Cys	Cys	Pro	Cys	Phe	Gly	Arg	Arg	Arg	Lys	210	215	220	
40	Asp	Lys	Lys	His	Ser	Lys	Asp	Gly	Gly	Asn	Ala	Asn	Gly	Leu	Ser	Leu	225	230	235	240
	Glu	Ala	Ala	Lys	Asp	Asp	Lys	Glu	Leu	Leu	Met	Ser	His	Met	Asn	Phe	245	250	255	
45	Glu	Lys	Lys	Phe	Gly	Gln	Ser	Ala	Ile	Phe	Val	Thr	Ser	Thr	Leu	Met	260	265	270	
50	Glu	Gln	Gly	Gly	Val	Pro	Pro	Ser	Ser	Ser	Pro	Ala	Ala	Leu	Leu	Lys	275	280	285	
	Glu	Ala	Ile	His	Val	Ile	Ser	Cys	Gly	Tyr	Glu	Asp	Lys	Thr	Glu	Trp	290	295	300	
55	Gly	Ser	Glu	Leu	Gly	Trp	Ile	Tyr	Gly	Ser	Ile	Thr	Glu	Asp	Ile	Leu				

305	310	315	320
Thr Gly Phe Lys Met His Cys Arg Gly Trp Arg Ser Ile Tyr Cys Met			
325	330	335	
Pro Lys Leu Pro Ala Phe Lys Gly Ser Ala Pro Ile Asn Leu Ser Asp			
340	345	350	
Arg Leu Asn Gln Val Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Phe			
355	360	365	
Phe Ser His His Cys Pro Ala Trp Tyr Gly Phe Lys Gly Gly Lys Leu			
370	375	380	
Lys Trp Leu Glu Arg Phe Ala Tyr Val Asn Thr Thr Ile Tyr Pro Phe			
385	390	395	400
Thr Ser Leu Pro Leu Leu Ala Tyr Cys Thr Leu Pro Ala Ile Cys Leu			
405	410	415	
Leu Thr Asp Lys Phe Ile Met Pro Pro Ile Ser Thr Phe Ala Ser Leu			
420	425	430	
Phe Phe Ile Ala Leu Phe Leu Ser Ile Phe Ala Thr Gly Ile Leu Glu			
435	440	445	
Leu Arg Trp Ser Gly Val Ser Ile Glu Glu Trp Trp Arg Asn Glu Gln			
450	455	460	
Phe Trp Val Ile Gly Gly Ile Ser Ala His Leu Phe Ala Val Ile Gln			
465	470	475	480
Gly Leu Leu Lys Val Leu Ala Gly Ile Asp Thr Asn Phe Thr Val Thr			
485	490	495	
Ser Lys Ala Thr Asp Asp Glu Glu Phe Gly Glu Leu Tyr Thr Phe Lys			
500	505	510	
Trp Thr Thr Leu Leu Ile Pro Pro Thr Thr Val Leu Ile Ile Asn Leu			
515	520	525	
Val Gly Val Val Ala Gly Ile Ser Asp Ala Ile Asn Asn Gly Tyr Gln			
530	535	540	
Ser Trp Gly Pro Leu Phe Gly Lys Leu Phe Phe Ser Phe Trp Val Ile			
545	550	555	560
Val His Leu Tyr Pro Phe Leu Lys Gly Leu Met Gly Arg Gln Asn Arg			
565	570	575	
Thr Pro Thr Ile Val Val Ile Trp Ser Val Leu Leu Ala Ser Ile Phe			
580	585	590	
Ser Leu Leu Trp Val Arg Ile Asp Pro Phe Val Met Lys Thr Lys Gly			
595	600	605	
Pro Asp Thr Thr Met Cys Gly Ile Asn Cys			
610	615		

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1086 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Gossypium hirsutum* L.

(C) INDIVIDUAL ISOLATE: Coker312

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 24..1086

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

	GGCAAGAGCT TTCATATCCT CCA ATG GAA GCC AGC GGC GGA CTC GTT GCG	50
	Met Glu Ala Ser Ala Gly Leu Val Ala	
	1 5	
	GGC TCT CAC AAC CGC AAT GAA CTT GTT GTC ATT CAT GGC CAT GAA GAG	98
	Gly Ser His Asn Arg Asn Glu Leu Val Val Ile His Gly His Glu Glu	
	10 15 20 25	
	OCT AAA OCT CTG AAG AAC TTG GAT GGT CAA GTT TGT GAG ATT TGT GGT	146
	Pro Lys Pro Leu Lys Asn Leu Asp Gly Gln Val Cys Glu Ile Cys Gly	
	30 35 40	
	GAT GAA ATT GGG TTG ACG GTC GAT GGA GAT CTT TTC GTG GCC TGC AAC	194
	Asp Glu Ile Gly Leu Thr Val Asp Gly Asp Leu Phe Val Ala Cys Asn	
	45 50 55	
	GAG TGT GGT TTT CCA GTT TGT AGG CCT TGT TAT GAG TAT GAA AGG AGA	242
	Glu Cys Gly Phe Pro Val Cys Arg Pro Cys Tyr Glu Tyr Glu Arg Arg	
	60 65 70	
	GAA GGG AGT CAA CAA TGT OCT CAA TGC AAA ACT AGA TAC AAG CGT CTC	290
	Glu Gly Ser Gln Gln Cys Pro Gln Cys Lys Thr Arg Tyr Lys Arg Leu	
	75 80 85	
	AAG GGG AGT CCG AGG GTG GAG GGA GAT GAA GAT GAA GAG GAT GTG GAT	338
	Lys Gly Ser Pro Arg Val Glu Gly Asp Glu Asp Glu Glu Asp Val Asp	
	90 95 100 105	
	GAT ATC GAA CAT GAA TTC AAC ATT GAT GAT GAA CAA AAC AAG TAT AGA	386
	Asp Ile Glu His Glu Phe Asn Ile Asp Asp Glu Gln Asn Lys Tyr Arg	
	110 115 120	
	AAT ATC GCT GAA TCG ATG CTT CAT GGA AAG ATG AGC TAC GGG AGA GGC	434
	Asn Ile Ala Glu Ser Met Leu His Gly Lys Met Ser Tyr Gly Arg Gly	
	125 130 135	
	OCT GAA GAC GAT GAA GGT TTG CAA ATC CCA CCG GGT TTA GCT GGT GTT	482

	Pro	Glu	Asp	Asp	Glu	Gly	Leu	Gln	Ile	Pro	Pro	Gly	Leu	Ala	Gly	Val	
					140			145					150				
5	CGA	TCT	CGG	COG	GTG	AGC	GGG	GAG	TTC	OCA	ATA	GGA	AGC	TCT	CTT	GCT	530
	Arg	Ser	Arg	Pro	Val	Ser	Gly	Glu	Phe	Pro	Ile	Gly	Ser	Ser	Leu	Ala	
					155			160					165				
	TAT	GGG	GAA	CAC	ATG	TCA	AAT	AAA	CGA	GTT	CAT	OCA	TAT	OCT	ATG	TCT	578
10	Tyr	Gly	Glu	His	Met	Ser	Asn	Lys	Arg	Val	His	Pro	Tyr	Pro	Met	Ser	
					170			175				180				185	
	GAA	OCT	GGA	AGT	GCA	AGA	TGG	GAT	GAA	AAG	AAA	GAG	GGA	GGA	TGG	AGA	626
	Glu	Pro	Gly	Ser	Ala	Arg	Trp	Asp	Glu	Lys	Lys	Glu	Gly	Gly	Trp	Arg	
					190					195					200		
15	GAA	AGG	ATG	GAT	GAT	TGG	AAA	ATG	CAG	CAA	GGG	AAT	TTG	GGT	OCT	GAA	674
	Glu	Arg	Met	Asp	Asp	Trp	Lys	Met	Gln	Gln	Gly	Asn	Leu	Gly	Pro	Glu	
					205				210					215			
20	OCT	GAT	GAT	GOC	TAT	GAT	GCT	GAC	ATG	GCT	ATG	CTT	GAT	GAA	GCT	AGG	722
	Pro	Asp	Asp	Ala	Tyr	Asp	Ala	Asp	Met	Ala	Met	Leu	Asp	Glu	Ala	Arg	
					220				225					230			
	CAG	OCA	TTG	TCA	AGG	AAA	GTG	OCA	ATT	GCA	TCG	AGC	AAA	ATC	AAT	OCT	770
25	Gln	Pro	Leu	Ser	Arg	Lys	Val	Pro	Ile	Ala	Ser	Ser	Lys	Ile	Asn	Pro	
					235			240					245				
	TAT	CGT	ATG	GTG	ATT	GTG	GCT	CGT	CTA	GTT	ATC	CTT	GCT	TTC	TTT	CTT	818
	Tyr	Arg	Met	Val	Ile	Val	Ala	Arg	Leu	Val	Ile	Leu	Ala	Phe	Phe	Leu	
					250			255				260				265	
30	CGC	TAT	CGG	ATT	TTG	AAC	COG	GTA	CAT	GAT	GCA	ATT	GGG	CTT	TGG	CTA	866
	Arg	Tyr	Arg	Ile	Leu	Asn	Pro	Val	His	Asp	Ala	Ile	Gly	Leu	Trp	Leu	
					270					275					280		
35	ACT	TCT	GTG	ATC	TGT	GAA	ATC	TGG	TTT	GOC	TTT	TCA	TGG	ATC	CTT	GAT	914
	Thr	Ser	Val	Ile	Cys	Glu	Ile	Trp	Phe	Ala	Phe	Ser	Trp	Ile	Leu	Asp	
					285				290					295			
	CAG	TTC	OCT	AAA	TGG	TTC	OCT	ATT	GAC	OGC	GAG	ACG	TAT	CTC	GAT	CGC	962
40	Gln	Phe	Pro	Lys	Trp	Phe	Pro	Ile	Asp	Arg	Glu	Thr	Tyr	Leu	Asp	Arg	
					300			305					310				
	CTT	TCC	CTC	AGG	TAT	GAG	AGG	GAA	GGT	GAG	CCC	AAC	ATG	CTT	GCT	TCT	1010
	Leu	Ser	Leu	Arg	Tyr	Glu	Arg	Glu	Gly	Glu	Pro	Asn	Met	Leu	Ala	Ser	
					315			320				325					
45	GTT	GAT	ATT	TTT	GTC	AGT	ACA	GTG	GAT	OCA	TTG	AAG	GGA	OCT	OCT	CTA	1058
	Val	Asp	Ile	Phe	Val	Ser	Thr	Val	Asp	Pro	Leu	Lys	Gly	Pro	Pro	Leu	
					330			335			340					345	
50	GTA	ACA	GCG	AAT	ACA	GTT	CTA	TCG	ATC	T							1086
	Val	Thr	Ala	Asn	Thr	Val	Leu	Ser	Ile								
					350												

(2) INFORMATION FOR SEQ ID NO: 8:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 354 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: N-terminal fragment

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

Met Glu Ala Ser Ala Gly Leu Val Ala Gly Ser His Asn Arg Asn Glu
 1 5 10 15
 Leu Val Val Ile His Gly His Glu Glu Pro Lys Pro Leu Lys Asn Leu
 20 25 30
 Asp Gly Gln Val Cys Glu Ile Cys Gly Asp Glu Ile Gly Leu Thr Val
 35 40 45
 Asp Gly Asp Leu Phe Val Ala Cys Asn Glu Cys Gly Phe Pro Val Cys
 50 55 60
 Arg Pro Cys Tyr Glu Tyr Glu Arg Arg Glu Gly Ser Gln Gln Cys Pro
 65 70 75 80
 Gln Cys Lys Thr Arg Tyr Lys Arg Leu Lys Gly Ser Pro Arg Val Glu
 85 90 95
 Gly Asp Glu Asp Glu Glu Asp Val Asp Asp Ile Glu His Glu Phe Asn
 100 105 110
 Ile Asp Asp Glu Gln Asn Lys Tyr Arg Asn Ile Ala Glu Ser Met Leu
 115 120 125
 His Gly Lys Met Ser Tyr Gly Arg Gly Pro Glu Asp Asp Glu Gly Leu
 130 135 140
 Gln Ile Pro Pro Gly Leu Ala Gly Val Arg Ser Arg Pro Val Ser Gly
 145 150 155 160
 Glu Phe Pro Ile Gly Ser Ser Leu Ala Tyr Gly Glu His Met Ser Asn
 165 170 175
 Lys Arg Val His Pro Tyr Pro Met Ser Glu Pro Gly Ser Ala Arg Trp
 180 185 190
 Asp Glu Lys Lys Glu Gly Gly Trp Arg Glu Arg Met Asp Asp Trp Lys
 195 200 205
 Met Gln Gln Gly Asn Leu Gly Pro Glu Pro Asp Asp Ala Tyr Asp Ala
 210 215 220
 Asp Met Ala Met Leu Asp Glu Ala Arg Gln Pro Leu Ser Arg Lys Val
 225 230 235 240
 Pro Ile Ala Ser Ser Lys Ile Asn Pro Tyr Arg Met Val Ile Val Ala
 245 250 255

Arg Leu Val Ile Leu Ala Phe Phe Leu Arg Tyr Arg Ile Leu Asn Pro
 260 265 270
 Val His Asp Ala Ile Gly Leu Trp Leu Thr Ser Val Ile Cys Glu Ile
 275 280 285
 Trp Phe Ala Phe Ser Trp Ile Leu Asp Gln Phe Pro Lys Trp Phe Pro
 290 295 300
 Ile Asp Arg Glu Thr Tyr Leu Asp Arg Leu Ser Leu Arg Tyr Glu Arg
 305 310 315 320
 Glu Gly Glu Pro Asn Met Leu Ala Ser Val Asp Ile Phe Val Ser Thr
 325 330 335
 Val Asp Pro Leu Lys Gly Pro Pro Leu Val Thr Ala Asn Thr Val Leu
 340 345 350
 Ser Ile

(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1000 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Gossypium hirsutum* L.
- (C) INDIVIDUAL ISOLATE: Coker312

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1000

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

GAC AAA GTC OGG CCG ACA TTC GTG AAG GAG CGT CGA GCT ATG AAG AGA	48
Asp Lys Val Arg Pro Thr Phe Val Lys Glu Arg Arg Ala Met Lys Arg	
1 5 10 15	
GAA TAT GAA GAA TTC AAG GTT AGG ATA AAT GCA CTT GTA GGC AAA GGC	96
Glu Tyr Glu Glu Phe Lys Val Arg Ile Asn Ala Leu Val Ala Lys Ala	
20 25 30	
CAA AAG GTT OCT CCA GAA GGG TGG ATC ATG CAA GAT GGG ACA CCA TGG	144
Gln Lys Val Pro Pro Glu Gly Trp Ile Met Gln Asp Gly Thr Pro Trp	
35 40 45	
OCA GGA AAC AAT ACT AAA GAT CAC CCT GGT ATG ATT CAA GTA TTT CTC	192
Pro Gly Asn Asn Thr Lys Asp His Pro Gly Met Ile Gln Val Phe Leu	
50 55 60	

	GGT CAA AGT GGA GGC CAT GAT AOC GAA GGA AAT GAG CTT OCT OGT CTC	240
	Gly Gln Ser Gly Gly His Asp Thr Glu Gly Asn Glu Leu Pro Arg Leu	
5	65 70 75 80	
	GTC TAT GTA TCT OGA GAG AAA AGG OCA GGT TTC TTG CAT CAC AAG AAA	288
	Val Tyr Val Ser Arg Glu Lys Arg Pro Gly Phe Leu His His Lys Lys	
	85 90 95	
10	GCT GGT GOC ATG AAC GOC CTT GTT OGT GTC TOG GGG GTG CTT ACA AAT	336
	Ala Gly Ala Met Asn Ala Leu Val Arg Val Ser Gly Val Leu Thr Asn	
	100 105 110	
	GCT OCT TTT ATG TTG AAC TTG GAT TGT GAC CAC TAT TTA AAT AAC AGC	384
15	Ala Pro Phe Met Leu Asn Leu Asp Cys Asp His Tyr Leu Asn Asn Ser	
	115 120 125	
	AAG GCT GTA AGA GAG GCT ATG TGT TTC TTG ATG GAC OCT CAA ATT GGA	432
	Lys Ala Val Arg Glu Ala Met Cys Phe Leu Met Asp Pro Gln Ile Gly	
	130 135 140	
20	AGA AAG GTT TGC TAT GTC CAA TTC OCT CAA OGT TTC GAT GGT ATT GAT	480
	Arg Lys Val Cys Tyr Val Gln Phe Pro Gln Arg Phe Asp Gly Ile Asp	
	145 150 155 160	
	AGA CAT GAT OGA TAT GOC AAT OGG AAC ACA GTT TTC TTT GAT ATT AAC	528
25	Arg His Asp Arg Tyr Ala Asn Arg Asn Thr Val Phe Phe Asp Ile Asn	
	165 170 175	
	ATG AAA GGT CTA GAT GGT ATA CAA GGC OCT GTA TAT GTC GGC ACG GGG	576
	Met Lys Gly Leu Asp Gly Ile Gln Gly Pro Val Tyr Val Gly Thr Gly	
30	180 185 190	
	TGT GTT TTC AGA AGG CAA GCT CTT TAT GGT TAT GAA OCT OCA AAG GGA	624
	Cys Val Phe Arg Arg Gln Ala Leu Tyr Gly Tyr Glu Pro Pro Lys Gly	
	195 200 205	
35	OCT AAG OGC OCG AAA ATG GTA AOC TGT GGT TGC TGC OCT TGC TTT GGA	672
	Pro Lys Arg Pro Lys Met Val Thr Cys Gly Cys Cys Pro Cys Phe Gly	
	210 215 220	
	OGC OGC AGA AAG GAC AAA AAG CAC TCT AAG GAT GGT GGA AAT GCA AAT	720
40	Arg Arg Arg Lys Asp Lys Lys His Ser Lys Asp Gly Gly Asn Ala Asn	
	225 230 235 240	
	GGT CTA AGC CTA GAA GCA GOC GAA GAT GAC AAG GAG TTA TTG ATG TOC	768
	Gly Leu Ser Leu Glu Ala Ala Glu Asp Asp Lys Glu Leu Leu Met Ser	
	245 250 255	
45	CAC ATG AAC TTT GAA AAG AAA TTT GGA CAA TCA GOC ATT TTT GTA ACT	816
	His Met Asn Phe Glu Lys Lys Phe Gly Gln Ser Ala Ile Phe Val Thr	
	260 265 270	
50	TCA ACA CTG ATG GAA CAA GGT GGT GTC OCT OCT TCT TCA AGC OCT GCA	864
	Ser Thr Leu Met Glu Gln Gly Gly Val Pro Pro Ser Ser Ser Pro Ala	

275 280 285
 GCT TTG CTC AAA GAA GCC ATT CAT GTA ATT AGT TGT GGT TAT GAA GAC 912
 Ala Leu Leu Lys Glu Ala Ile His Val Ile Ser Cys Gly Tyr Glu Asp
 290 295 300
 AAA ACC GAA TGG GGA AGC GAG CTT GGC TGG ATT TAC GGC TCG ATT ACA 960
 Lys Thr Glu Trp Gly Ser Glu Leu Gly Trp Ile Tyr Gly Ser Ile Thr
 305 310 315 320
 GAA GAT ATC TTA ACA GGT TTC AAG ATG CAT TGC CGT GGA T 1000
 Glu Asp Ile Leu Thr Gly Phe Lys Met His Cys Arg Gly
 325 330

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 333 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: internal fragment

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

Asp Lys Val Arg Pro Thr Phe Val Lys Glu Arg Arg Ala Met Lys Arg
 1 5 10 15
 Glu Tyr Glu Glu Phe Lys Val Arg Ile Asn Ala Leu Val Ala Lys Ala
 20 25 30
 Gln Lys Val Pro Pro Glu Gly Trp Ile Met Gln Asp Gly Thr Pro Trp
 35 40 45
 Pro Gly Asn Asn Thr Lys Asp His Pro Gly Met Ile Gln Val Phe Leu
 50 55 60
 Gly Gln Ser Gly Gly His Asp Thr Glu Gly Asn Glu Leu Pro Arg Leu
 65 70 75 80
 Val Tyr Val Ser Arg Glu Lys Arg Pro Gly Phe Leu His His Lys Lys
 85 90 95
 Ala Gly Ala Met Asn Ala Leu Val Arg Val Ser Gly Val Leu Thr Asn
 100 105 110
 Ala Pro Phe Met Leu Asn Leu Asp Cys Asp His Tyr Leu Asn Asn Ser
 115 120 125
 Lys Ala Val Arg Glu Ala Met Cys Phe Leu Met Asp Pro Gln Ile Gly
 130 135 140
 Arg Lys Val Cys Tyr Val Gln Phe Pro Gln Arg Phe Asp Gly Ile Asp
 145 150 155 160
 Arg His Asp Arg Tyr Ala Asn Arg Asn Thr Val Phe Phe Asp Ile Asn
 165 170 175

Met Lys Gly Leu Asp Gly Ile Gln Gly Pro Val Tyr Val Gly Thr Gly
 180 185 190
 5 Cys Val Phe Arg Arg Gln Ala Leu Tyr Gly Tyr Glu Pro Pro Lys Gly
 195 200 205
 Pro Lys Arg Pro Lys Met Val Thr Cys Gly Cys Cys Pro Cys Phe Gly
 210 215 220
 10 Arg Arg Arg Lys Asp Lys Lys His Ser Lys Asp Gly Gly Asn Ala Asn
 225 230 235 240
 Gly Leu Ser Leu Glu Ala Ala Glu Asp Asp Lys Glu Leu Leu Met Ser
 245 250 255
 15 His Met Asn Phe Glu Lys Lys Phe Gly Gln Ser Ala Ile Phe Val Thr
 260 265 270
 Ser Thr Leu Met Glu Gln Gly Gly Val Pro Pro Ser Ser Ser Pro Ala
 275 280 285
 20 Ala Leu Leu Lys Glu Ala Ile His Val Ile Ser Cys Gly Tyr Glu Asp
 290 295 300
 Lys Thr Glu Trp Gly Ser Glu Leu Gly Trp Ile Tyr Gly Ser Ile Thr
 305 310 315 320
 25 Glu Asp Ile Leu Thr Gly Phe Lys Met His Cys Arg Gly
 325 330

(2) INFORMATION FOR SEQ ID NO: 11:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 622 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) FRAGMENT TYPE: C-terminal fragment

(ix) FEATURE:

(A) NAME/KEY:

(B) LOCATION:

(D) OTHER INFORMATION: Xaa indicates Glu or Lys

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

45 Asp Lys Val Arg Pro Thr Phe Val Lys Glu Arg Arg Ala Met Lys Arg
 1 5 10 15
 Glu Tyr Glu Glu Phe Lys Val Arg Ile Asn Ala Leu Val Ala Lys Ala
 20 25 30
 50 Gln Lys Val Pro Pro Glu Gly Trp Ile Met Gln Asp Gly Thr Pro Trp
 35 40 45
 Pro Gly Asn Asn Thr Lys Asp His Pro Gly Met Ile Gln Val Phe Leu
 50 55 60
 55

Gly Gln Ser Gly Gly His Asp Thr Glu Gly Asn Glu Leu Pro Arg Leu
 65 70 75 80
 5 Val Tyr Val Ser Arg Glu Lys Arg Pro Gly Phe Leu His His Lys Lys
 85 90 95
 Ala Gly Ala Met Asn Ala Leu Val Arg Val Ser Gly Val Leu Thr Asn
 100 105 110
 10 Ala Pro Phe Met Leu Asn Leu Asp Cys Asp His Tyr Leu Asn Asn Ser
 115 120 125
 Lys Ala Val Arg Glu Ala Met Cys Phe Leu Met Asp Pro Gln Ile Gly
 130 135 140
 15 Arg Lys Val Cys Tyr Val Gln Phe Pro Gln Arg Phe Asp Gly Ile Asp
 145 150 155 160
 Arg His Asp Arg Tyr Ala Asn Arg Asn Thr Val Phe Phe Asp Ile Asn
 165 170 175
 20 Met Lys Gly Leu Asp Gly Ile Gln Gly Pro Val Tyr Val Gly Thr Gly
 180 185 190
 Cys Val Phe Arg Arg Gln Ala Leu Tyr Gly Tyr Glu Pro Pro Lys Gly
 195 200 205
 25 Pro Lys Arg Pro Lys Met Val Thr Cys Gly Cys Cys Pro Cys Phe Gly
 210 215 220
 Arg Arg Arg Lys Asp Lys Lys His Ser Lys Asp Gly Gly Asn Ala Asn
 225 230 235 240
 30 Gly Leu Ser Leu Glu Ala Ala Xaa Asp Asp Lys Glu Leu Leu Met Ser
 245 250 255
 His Met Asn Phe Glu Lys Lys Phe Gly Gln Ser Ala Ile Phe Val Thr
 260 265 270
 35 Ser Thr Leu Met Glu Gln Gly Gly Val Pro Pro Ser Ser Ser Pro Ala
 275 280 285
 Ala Leu Leu Lys Glu Ala Ile His Val Ile Ser Cys Gly Tyr Glu Asp
 290 295 300
 40 Lys Thr Glu Trp Gly Ser Glu Leu Gly Trp Ile Tyr Gly Ser Ile Thr
 305 310 315 320
 Glu Asp Ile Leu Thr Gly Phe Lys Met His Cys Arg Gly Trp Arg Ser
 325 330 335
 45 Ile Tyr Cys Met Pro Lys Leu Pro Ala Phe Lys Gly Ser Ala Pro Ile
 340 345 350
 Asn Leu Ser Asp Arg Leu Asn Gln Val Leu Arg Trp Ala Leu Gly Ser
 355 360 365
 50 Val Glu Ile Phe Phe Ser His His Cys Pro Ala Trp Tyr Gly Phe Lys
 370 375 380
 55 Gly Gly Lys Leu Lys Trp Leu Glu Arg Phe Ala Tyr Val Asn Thr Thr

385					390					395					400
Ile	Tyr	Pro	Phe	Thr	Ser	Leu	Pro	Leu	Leu	Ala	Tyr	Cys	Thr	Leu	Pro
				405					410					415	
Ala	Ile	Cys	Leu	Leu	Thr	Asp	Lys	Phe	Il	Met	Pro	Pro	Ile	Ser	Thr
			420					425						430	
Phe	Ala	Ser	Leu	Phe	Phe	Ile	Ala	Leu	Phe	Leu	Ser	Ile	Phe	Ala	Thr
		435					440					445			
Gly	Ile	Leu	Glu	Leu	Arg	Trp	Ser	Gly	Val	Ser	Ile	Glu	Glu	Trp	Trp
	450					455					460				
Arg	Asn	Glu	Gln	Phe	Trp	Val	Ile	Gly	Gly	Ile	Ser	Ala	His	Leu	Phe
465					470					475					480
Ala	Val	Ile	Gln	Gly	Leu	Leu	Lys	Val	Leu	Ala	Gly	Ile	Asp	Thr	Asn
			485						490					495	
Phe	Thr	Val	Thr	Ser	Lys	Ala	Thr	Asp	Asp	Glu	Glu	Phe	Gly	Glu	Leu
			500					505					510		
Tyr	Thr	Phe	Lys	Trp	Thr	Thr	Leu	Leu	Ile	Pro	Pro	Thr	Thr	Val	Leu
	515						520					525			
Ile	Ile	Asn	Leu	Val	Gly	Val	Val	Ala	Gly	Ile	Ser	Asp	Ala	Ile	Asn
	530					535					540				
Asn	Gly	Tyr	Gln	Ser	Trp	Gly	Pro	Leu	Phe	Gly	Lys	Leu	Phe	Phe	Ser
545					550					555					560
Phe	Trp	Val	Ile	Val	His	Leu	Tyr	Pro	Phe	Leu	Lys	Gly	Leu	Met	Gly
			565					570						575	
Arg	Gln	Asn	Arg	Thr	Pro	Thr	Ile	Val	Val	Ile	Trp	Ser	Val	Leu	Leu
			580					585						590	
Ala	Ser	Ile	Phe	Ser	Leu	Leu	Trp	Val	Arg	Ile	Asp	Pro	Phe	Val	Met
		595					600				605				
Lys	Thr	Lys	Gly	Pro	Asp	Thr	Thr	Met	Cys	Gly	Ile	Asn	Cys		
610						615					620				

(2) INFORMATION FOR SEQ ID NO: 12:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 9 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(v) **FRAGMENT TYPE:** internal fragment

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

Gln Xaa Xaa Xaa Xaa Xaa Xaa Arg Trp

1

5

(2) INFORMATION FOR SEQ ID NO: 13:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 50 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "Synthetic DNA"

(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

GAGAGAGAGA GAGAGAGAGA ACTAGTCTCG AGTTTTTTTT TTTTTTTTTT

50

(2) INFORMATION FOR SEQ ID NO: 14:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 13 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "Synthetic DNA"

(1x) FEATURE:

- (A) NAME/KEY:
- (B) LOCATION:1..4
- (D) OTHER INFORMATION: single strand

(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

AATTCGGCAC GAG

13

(2) INFORMATION FOR SEQ ID NO: 15:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(11) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "Synthetic DNA"

(x1) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

GACTGAAGAT AAGCCAAAAG

20

(2) INFORMATION FOR SEQ ID NO: 16:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 19 base pairs
- (B) TYPE: nucleic acid

(C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

5 (ii) MOLECULE TYPE: other nucleic acid
 (A) DESCRIPTION: /desc = "Synthetic DNA"
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:
 GGAATGATGA ATTTGCOGG 19

10 (2) INFORMATION FOR SEQ ID NO: 17:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 15 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: other nucleic acid
 (A) DESCRIPTION: /desc = "Synthetic DNA"
 20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:
 TGCAGGCAAC TTTGGCATGC 20

(2) INFORMATION FOR SEQ ID NO: 18:
 25 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 30 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 30 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: other nucleic acid
 (A) DESCRIPTION: /desc = "Synthetic DNA"
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:
 35 AGCAACAAGA GCAAGATGAG GAGGATGACT 30

(2) INFORMATION FOR SEQ ID NO: 19:
 (i) SEQUENCE CHARACTERISTICS:
 40 (A) LENGTH: 28 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 45 (ii) MOLECULE TYPE: other nucleic acid
 (A) DESCRIPTION: /desc = "Synthetic DNA"
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:
 COGGATCCTT CAACCTTCT TCGATTTC 28

50 (2) INFORMATION FOR SEQ ID NO: 20:

55

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 29 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "Synthetic DNA"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

COGGATOCAC GGCAATGCAT CTTGAAACC

29

(2) INFORMATION FOR SEQ ID NO: 21:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "Synthetic DNA"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

GGTTAGCATA TTGTTTGTAG CATTGGG

27

(2) INFORMATION FOR SEQ ID NO: 22:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "Synthetic DNA"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:

ATCAATGAAA TATGTATAGT TCATAGC

27

(2) INFORMATION FOR SEQ ID NO: 23:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

- (A) DESCRIPTION: /desc = "Synthetic DNA"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

CTTTOGTTCT TTTGGTTTIG OCATGGC

27

(2) INFORMATION FOR SEQ ID NO: 24:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "Synthetic DNA"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24:

AGACTTTTTA CAAACAAGAT AAATCOC

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Claims

1. A DNA coding for any one of the following proteins (A) to (C):

(A) a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO: 2 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 2;

(B) a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO: 4 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 4; and

(C) a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO: 8 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 8, and an amino acid sequence shown in SEQ ID NO: 11 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO: 11.

2. A recombinant vector comprising all or a part of the DNA as defined in claim 1.

3. A transformed cell transformed with the DNA as defined in claim 1.

4. A method for controlling cellulose synthesis in a cell, comprising the steps of introducing the DNA as defined in claim 1 into the cell, and expressing RNA having a nucleotide sequence homologous to the DNA as defined in claim 1 or a nucleotide sequence complementary to the DNA as defined in claim 1.

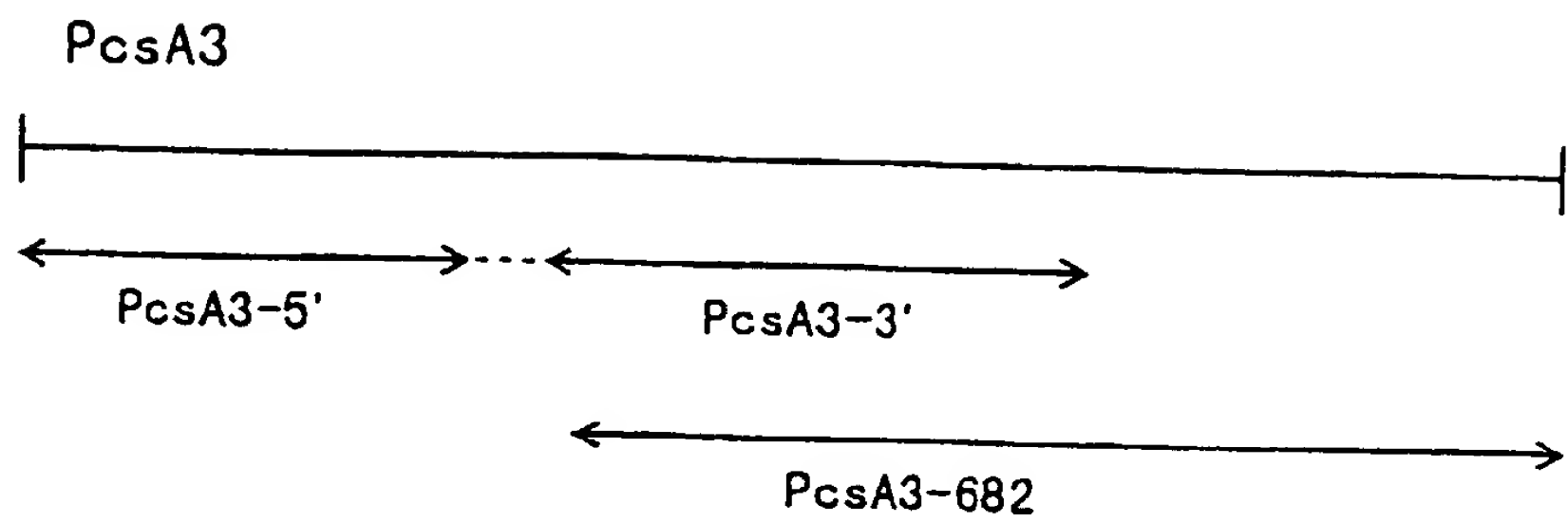


FIG. 1

SEQ ID NO: 14

5' AATTCGGCACGAG 3'
3' GCCGTGCTC 5' ---

FIG. 2

	10	20	30	40	50	60
PcsA3-682	CCGACATTCGTGAAGGAGCGTCGAGCTATGAAGAGAGAATATGAAGAATTCAAGGTTAGG					
(SEQ ID NO: 5)					
PcsA3-3'	CCGACATTCGTGAAGGAGCGTCGAGCTATGAAGAGAGAATATGAAGAATTCAAGGTTAGG					
(SEQ ID NO: 9)	20	30	40	50	60	70
	70	80	90	100	110	120
PcsA3-682	ATAAATGCACTTGTAGCCAAAGCCCCAAAAGGTTCCCTCCAGAAGGGTGGATCATGCAAGAT					
					
PcsA3-3'	ATAAATGCACTTGTAGCCAAAGCCCCAAAAGGTTCCCTCCAGAAGGGTGGATCATGCAAGAT					
	80	90	100	110	120	130
	130	140	150	160	170	180
PcsA3-682	GGGACACCATGGCCAGGAAACAATACTAAAGATCACCTGGTATGATTCAAGTATTCTC					
					
PcsA3-3'	GGGACACCATGGCCAGGAAACAATACTAAAGATCACCTGGTATGATTCAAGTATTCTC					
	140	150	160	170	180	190
	190	200	210	220	230	240
PcsA3-682	GGTCAAAGTGGAGGCCATGATACCGAAGGAAATGAGCTTCCTCGTCTCGTCTATGTATCT					
					
PcsA3-3'	GGTCAAAGTGGAGGCCATGATACCGAAGGAAATGAGCTTCCTCGTCTCGTCTATGTATCT					
	200	210	220	230	240	250
	250	260	270	280	290	300
PcsA3-682	CGAGAGAAAAGGCCTGGTTTCTTGCATCACAAGAAAGCTGGTGCCATGAACGCCCTTGTT					
*					
PcsA3-3'	CGAGAGAAAAGGCCTGGTTTCTTGCATCACAAGAAAGCTGGTGCCATGAACGCCCTTGTT					
	260	270	280	290	300	310
	310	320	330	340	350	360
PcsA3-682	CGGGTCTCGGGGTGCTCACAAATGCTCCTTTTATGTTGAAGTTGGATTGTGACCATTAT					
	:*:.....*:.....*					
PcsA3-3'	CGTGTCTCGGGGTGCTTACAAATGCTCCTTTTATGTTGAAGTTGGATTGTGACCACTAT					
	320	330	340	350	360	370
	370	380	390	400	410	420
PcsA3-682	TTAAATAACAGCAAGGCTGTAAGAGAGGCTATGTGTTTCTTGATGGACCCTCAAATTGGA					
					
PcsA3-3'	TTAAATAACAGCAAGGCTGTAAGAGAGGCTATGTGTTTCTTGATGGACCCTCAAATTGGA					
	380	390	400	410	420	430
	430	440	450	460	470	480
PcsA3-682	AGAAAGGTTTGCTATGTCCAATTCCTCAACGTTTCGATGGTATTGATAGACATGATCGA					
					
PcsA3-3'	AGAAAGGTTTGCTATGTCCAATTCCTCAACGTTTCGATGGTATTGATAGACATGATCGA					
	440	450	460	470	480	490
	490	500	510	520	530	540
PcsA3-682	TATGCCAATCGGAACACAGTTTTCTTTGATATTAACATGAAAGGTCTAGATGGTATACAA					
					
PcsA3-3'	TATGCCAATCGGAACACAGTTTTCTTTGATATTAACATGAAAGGTCTAGATGGTATACAA					
	500	510	520	530	540	550

FIG. 3

	550	560	570	580	590	600
PcsA3-682	GGCCCTGTATATGTGGGCACGGGGTGTGTTTTCAGAAGGCAAGCTCTTTATGGTTATGAA					
(SEQ ID NO: 5)					
PcsA3-3'	GGCCCTGTATATGTGGGCACGGGGTGTGTTTTCAGAAGGCAAGCTCTTTATGGTTATGAA					
(SEQ ID NO: 9)	560	570	580	590	600	610
	610	620	630	640	650	660
PcsA3-682	CCTCCAAAGGGACCTAAGCGCCCGAAAATGGTAACCTGTGGTTGCTGCCCTTGTTTTGGA					
*					
PcsA3-3'	CCTCCAAAGGGACCTAAGCGCCCGAAAATGGTAACCTGTGGTTGCTGCCCTTGCTTTGGA					
	620	630	640	650	660	670
	670	680	690	700	710	720
PcsA3-682	CGCCGCAGAAAGGACAAAAAGCACTCTAAGGATGGTGGAAATGCAAATGGTCTAAGCCTA					
					
PcsA3-3'	CGCCGCAGAAAGGACAAAAAGCACTCTAAGGATGGTGGAAATGCAAATGGTCTAAGCCTA					
	680	690	700	710	720	730
	730	740	750	760	770	780
PcsA3-682	GAAGCAGCCAAAGATGACAAGGAGTTATTGATGTCCACATGAACCTTGAAAAGAAATTT					
*					
PcsA3-3'	GAAGCAGCCGAAGATGACAAGGAGTTATTGATGTCCACATGAACCTTGAAAAGAAATTT					
	740	750	760	770	780	790
	790	800	810	820	830	840
PcsA3-682	GGACAATCAGCCATTTTGTAACTTCAACACTGATGGAACAAGGTGGTGTCCCTCCTTCT					
					
PcsA3-3'	GGACAATCAGCCATTTTGTAACTTCAACACTGATGGAACAAGGTGGTGTCCCTCCTTCT					
	800	810	820	830	840	850
	850	860	870	880	890	900
PcsA3-682	TCAAGCCCCGCAGCTTTGCTCAAAGAAGCCATTGATGTAATTAGTTGTGGTTATGAAGAC					
*					
PcsA3-3'	TCAAGCCCTGCAGCTTTGCTCAAAGAAGCCATTGATGTAATTAGTTGTGGTTATGAAGAC					
	860	870	880	890	900	910
	910	920	930	940	950	960
PcsA3-682	AAAACAGAATGGGGAAGCGAGCTTGGCTGGATTTACGGCTCGATTACAGAAGATATCTTA					
*					
PcsA3-3'	AAAACCGAATGGGGAAGCGAGCTTGGCTGGATTTACGGCTCGATTACAGAAGATATCTTA					
	920	930	940	950	960	970
	970	980				
PcsA3-682	ACAGGATTCAAGATGCATTGCCGTGGAT					
*					
PcsA3-3'	ACAGGTTTCAAGATGCATTGCCGTGGAT					
	980	990	1000			

FIG. 4

(19)



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(11)

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(12)

EUROPEAN PATENT APPLICATION

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(54) **Cellulose synthase gene**

(57) mRNA was extracted at the stage for cotton plant fibrous cells to accumulate cellulose, and cDNA's complementary thereto were synthesized to construct a cDNA library. Clones of a number of 750 were arbitrarily selected from the library, and they were randomly subjected from to sequencing. Those having homology to

an amino acid sequence deduced from a gene of cellulose 4- β -glucosyltransferase (bcsA) of cellulose synthase operon of acetic acid bacterium were selected from obtained nucleotide sequences of the respective clones. Thus, DNA coding for cellulose synthase was obtained.

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European Patent
Office

EUROPEAN SEARCH REPORT

Application Number
EP 98 30 2489

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.6)
P,X	WO 98 00549 A (THE AUSTRALIAN NATIONAL UNIVERSITY; COMMONWEALTH SCIENTIFIC...) 8 January 1998 * page 1, line 3 - line 11 * * page 2, line 21 - page 7, line 28 * * example 8 * 'Sequence Listing: SEQ ID NO.9 and 10' ---	1-4	C12N15/54 C12N9/10
X,D	PEAR, J.R. ETAL.: "Higher plants contain homologs of the bacterial celA genes encoding the catalytic subunit of cellulose synthase" PROC.NATL.ACAD.SCI.USA, vol. 93, October 1996, pages 12637-12642, XP002061424 * the whole document * ---	1-4	
Y	WO 91 13988 A (THE BOARD OF REGENTS, THE UNIVERSITY OF TEXAS SYSTEM) 19 September 1991 * page 1, line 18 - line 26 * * page 5, line 15 - page 8, line 13 * * figure 1; examples I,II,IV,V * ---	1-4	TECHNICAL FIELDS SEARCHED (Int.Cl.6) C12N
Y	LI, L. ET AL.: "β-Glucan synthesis in the cotton fiber" PLANT PHYSIOLOGY, vol. 101, no. 4, 1993, pages 1149-1156, XP002087180 * page 1149 * * page 1154 - page 1155 * 'Abstract' and 'Discussion' ---	1-4	
E	WO 98 18949 A (CALGENE, INC.) 7 May 1998 * page 7, line 14 - page 9, line 25 * * figures 3,6,8; examples 1-7 * -----	1-4	
The present search report has been drawn up for all claims			
Place of search MUNICH		Date of completion of the search 8 December 1998	Examiner Donath, C
CATEGORY OF CITED DOCUMENTS X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document			

EP 0 875 575 A3 (P04C01)



European Patent
Office

Application Number
EP 98 30 2489

CLAIMS INCURRING FEES

The present European patent application comprised at the time of filing more than ten claims.

- ☐ Only part of the claims have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims and for those claims for which claims fees have been paid, namely claim(s):
- ☐ No claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims.

LACK OF UNITY OF INVENTION

The Search Division considers that the present European patent application does not comply with the requirements of unity of invention and relates to several inventions or groups of inventions, namely:

see sheet 8

- ☐ All further search fees have been paid within the fixed time limit. The present European search report has been drawn up for all claims.
- ☐ As all searchable claims could be searched without effort justifying an additional fee, the Search Division did not invite payment of any additional fee.
- ☐ Only part of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the inventions in respect of which search fees have been paid, namely claims:
- ☒ None of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the invention first mentioned in the claims, namely claims:
- 1-4 (partially)



European Patent
Office

LACK OF UNITY OF INVENTION
SHEET B

Application Number

EP 98 30 2489

The Search Division considers that the present European patent application does not comply with the requirements of unity of invention and relates to several inventions or groups of inventions, namely:

1. Claims: 1-4 (partially)

Claims 1- 4 (partially) refer to a DNA coding for a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO:2 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO:2, a recombinant vector comprising all or part of said DNA, a cell being transformed with said DNA, and a method for controlling cellulose synthesis in a cell by the use of said DNA.

2. Claims: 1-4 (partially)

Claims 1- 4 (partially) refer to a DNA coding for a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO:4 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO:4, a recombinant vector comprising all or part of said DNA, a cell being transformed with said DNA, and a method for controlling cellulose synthesis in a cell by the use of said DNA.

3. Claims: 1-4 (partially)

Claims 1- 4 (partially) refer to a DNA coding for a protein having a cellulose synthase activity and comprising an amino acid sequence shown in SEQ ID NO:8 and in SEQ ID NO:11 or an amino acid sequence involving deletion, substitution, insertion, or addition of one or several amino acids relevant to SEQ ID NO:8 and/or SEQ ID NO:11, a recombinant vector comprising all or part of said DNA, a cell being transformed with said DNA, and a method for controlling cellulose synthesis in a cell by the use of said DNA.